

21 Preventive and Therapeutic Effects of Plant Polyphenols through Suppression of Nuclear Factor-Kappa B

Navindra P. Seeram
UCLA, Los Angeles

*Haruyo Ichikawa, Shishir Shishodia, and
Bharat B. Aggarwal*
University of Texas, Houston

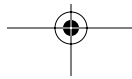


CONTENTS

21.1 Overview.....	243
21.2 Introduction.....	244
21.3 NF-κB and Disease	244
21.4 NF-κB as a Therapeutic Target.....	245
Acknowledgments	262
Abbreviations	263
References	264

21.1 OVERVIEW

Nuclear transcription factor-κB (NF-κB) regulates the expression of over 200 different genes. The activation of NF-κB has now been linked with a variety of inflammatory diseases, including cancer, atherosclerosis, myocardial infarction, diabetes, allergy, asthma, arthritis, Crohn’s disease, multiple sclerosis, Alzheimer’s disease, osteoporosis, psoriasis, septic shock, and AIDS. There is much evidence suggesting that phytochemicals can inhibit the pathways that lead to the activation of this transcription factor and have the potential to prevent and treat





the diseases mentioned. These phytochemicals are derived from plants such as turmeric, red pepper, cloves, ginger, cumin, anise and fennel, rosemary, garlic, green tea, basil, cauliflower, cabbage, artichoke, lemon, and pomegranate.

21.2 INTRODUCTION

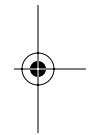
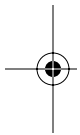
Plant extracts and natural compounds purified from plants have been used by humans for many centuries for the treatment and alleviation of a variety of inflammation-related diseases, including cancer. Eastern medicine, viz., traditional Chinese medicine (TCM) and the Indian ayurvedic system of medicine, continue to prescribe complex mixtures of herbs and herbal extracts for the treatment of cancer. Recent research has shown that a mechanism-based approach that targets the means by which cancer cells prosper has significant advantages over the current methods of cancer treatment, including chemotherapy, with their attendant adverse **effects**. The regulation of the cell cycle (cell survival, proliferation, and death) requires the integration of a myriad of cell-signaling factors, including those that direct the transcription of genes coding for integral cell proteins. Nuclear factor (NF)- κ B is a transcription factor that regulates the expression of genes involved in cancer and other diseases.

AU: Change
OK?

21.3 NF- κ B AND DISEASE

NF- κ B, discovered by David Baltimore in 1986, is a ubiquitous factor that resides in the cytoplasm in an inactive state. When activated, it is translocated to the nucleus and induces gene transcription. NF- κ B is activated by free radicals, inflammatory stimuli, carcinogens, tumor promoters, endotoxins, gamma radiation, UV light, and x-rays. On activation, it induces the expression of more than 200 genes, and these genes have been shown to suppress apoptosis and induce cellular transformation, proliferation, invasion, metastasis, chemoresistance, radioresistance, and inflammation.¹⁻³ The activated form of NF- κ B has been found to mediate cancer,^{1,4,5} atherosclerosis,⁶ myocardial infarction,⁷ diabetes,⁸ allergy,^{9,10} asthma,¹¹ arthritis,¹² Crohn's disease,¹³ multiple sclerosis,¹⁴ Alzheimer's disease,^{15,16} osteoporosis, psoriasis, septic shock, AIDS, and other inflammatory diseases¹⁷⁻¹⁹ (Figure 21.1). That NF- κ B has been linked to wide variety of diseases is not too surprising because most diseases are caused by dysregulated inflammatory mechanisms.²⁰ Thus, agents that can suppress NF- κ B activation can, in principle, either prevent, delay the onset, or treat NF- κ B -linked diseases.

Ever since research has shown that there is an intrinsic link between inflammation and various diseases, it has become obvious that inhibition of NF- κ B activity is desirable in the treatment of not only inflammation but also the disease itself. For instance, aberrant NF- κ B activation is a known factor in oncogenesis, tumor growth, and metastasis, and specific constitutive activation of NF- κ B has been identified in a number of cancers including, breast, ovarian, colon, and prostate cancer and Hodgkin's lymphoma (for references see Reference 1). Hence,



AU: Chapter title needs to be reduced in running head to fit. Pls. check wording.

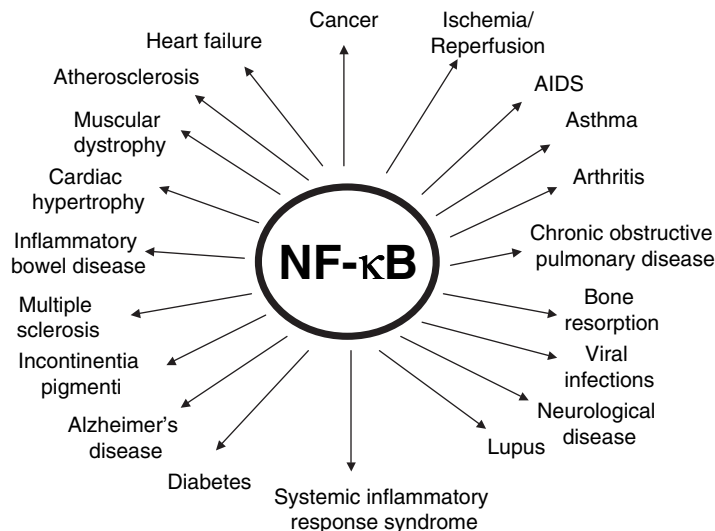


FIGURE 21.1 NF-κB-linked diseases.

this transcription factor is an important target in the prevention of diseases and their treatment in humans.

21.4 NF-κB AS A THERAPEUTIC TARGET

NF-κB represents an important and very attractive therapeutic target for plant-derived polyphenols. Much attention has been paid in the last decade to the identification of compounds that selectively interfere with this pathway. More recently, a great number of plant-derived natural products have been evaluated as possible inhibitors of the NF-κB pathway (Table 21.1, Figure 21.2). This chapter focuses on plant extracts, plant isolates, and distinct classes of plant-derived compounds that form part of this group. It is noteworthy that there are also reports of synthetic compounds and compounds from nonplant sources (e.g., caffeic acid phenethyl ester [CAPE] identified from honey bee propolis) that are known to block the activity of NF-κB²¹.

NF-κB plays a central role in inflammation, and research has made it clear that most diseases are linked to inflammation. Because NF-κB can also regulate the expression of many key genes involved in a variety of human cancers, it represents a relevant and promising target for new therapeutic agents. Many pharmaceutical and biotechnology companies have drug discovery programs that target NF-κB and have been investing heavily in the search for proteins that regulate this transcription factor. However, due to the ubiquitous nature of NF-κB, many of these drugs may exhibit undesirable side effects.

The mechanism-based approach to combat diseases from different angles with combinations of naturally derived compounds has a distinct advantage in

TABLE 21.1
Natural Products from Plants that Exhibit Chemopreventive and Therapeutic Activities against Cancer

Compound	Source	Botanical Name	Structure	↓NF-κB	P	T	Ref.
Polyphenol							
Amentoflavone (biapigenin)*	Ginkgo	<i>Ginkgo biloba</i> ; <i>Cypress</i> spp.; <i>Galeobdolon chinense</i> ; <i>Garcinia intermedia</i> ; <i>Selaginella</i> spp.; <i>Biophytum sensitivum</i>		+	—	—	22,23
Apigenin*	Fruits and vegetables	<i>Scutellaria</i> spp. (incl. in Chinese herbal mixture, PCSPES, <i>Huang-Qi</i> ; <i>Qingkailing</i> ; <i>Shuanghuanglian</i> etc.); <i>Cirisium</i> spp.; <i>Crotalaria</i> spp.; <i>Quercus nutgall</i> ; <i>Matricaria recutita</i> ; <i>Saussurea medusa</i> ; <i>Lantana montevidensis</i> Briq.	4',5,7-Trihydroxyflavone	+	—	—	24–26
Arctigenin* and demethylarctigenin		<i>Arctium lappa</i> ; <i>Centaurea</i> spp.; <i>Torreya nucifera</i>		+	—	—	27
Auraptene	Citrus fruits (hassaku, grapefruit, natudaidai)	<i>Citrus</i> spp.	7-Geranyl-oxy coumarin	—	—	—	28
Baicalein* and its derivatives ^a	Skullcap	<i>Scutellaria</i> spp. (included in Chinese herbal mixture, PCSPES, <i>Huang-Qi</i> ; <i>Qingkailing</i> ; <i>Shuanghuanglian</i> , etc.)	5,6,7-Trihydroxyflavone	+	+	+	29–31
Blueberry and berry mix	Blueberry, black currant, raspberry, strawberry	<i>Rubus</i> spp.; <i>Vaccinium</i> spp.; <i>Fragaria ananassa</i>		+	—	—	32–34
Cannabinol*	Hemp seed oil, marijuana	<i>Cannabis</i> spp.	6,6,9-Trimethyl-3-pentyl-6H-dibenzof[b,d]pyran-1-ol;	+	—	+	35–37

AU: What does this symbol indicate?

AU: Should this be "component of"?

AU: Should this be "and other berries"?



Preventive and Therapeutic Effects of Plant Polyphenols

247

Catalposide*	<i>Catalpa</i> spp.; <i>Veronica</i> spp.	Oxireno[4,5]cyclopenta [1,2-c]pyran- β -D- glucopyranoside	+	—	—	38
Catechins* (and theaflavins*) ^b	Green tea (including fermented, i.e., black teas), spotted knapweed, shea kernels, cocoa Basil, sage, rosemary	<i>Camellia sinensis</i> ; <i>Centaurea maculosa</i> ; <i>Vitellaria paradoxa</i> ; <i>Theobroma cacao</i>	+	+	+	39–41
Cirsimaritin*	Basil, sage, rosemary	<i>Cirsium maritimum</i> ; <i>Ocimum sanctum</i> ; <i>Salvia officinalis</i> ; <i>Rosmarinus officinalis</i>	—	—	—	42
Curcumin*	Turmeric (<i>haldi</i>)	<i>Curcuma longa</i>	+	+	+	—
Ellagic acid*	Strawberries, raspberries, blackberries, bayberries, fuejtoa, pomegranates, pineapple, walnuts Aloe vera	<i>Fragaria ananassa</i> ; <i>Rubus idaeus</i> ; <i>Punica granatum</i> ; <i>Juglans regia</i>	—	—	—	34,44
Emodin*	Aloe vera	<i>Polygonum</i> spp.; <i>Cassia</i> spp.; <i>Glossostemon brugueri</i> (<i>moghat</i>); <i>Rheum</i> spp. (<i>rhubarb</i>); <i>Hovenia acerba</i>	+	+	—	45
Flavopiridol ^c		5,7-Dihydroxy-8-(4- <i>N</i> -methyl- 2-hydroxypyridyl)-6'- chloroflavone hydrochloride 3-(4-Hydroxyphenyl)-5,7- dihydroxy-chromen-4-one	+	—	—	46
Genistein*	Soybeans, chickpea, kudzu root	<i>Glycine max</i> ; <i>Cicer arietinum</i> ; <i>Pueraria lobata radix</i> ; <i>Desmodium spp.</i> <i>Glossogyne tenuifolia</i>	+	+	+	47,48
<i>Glossogyne tenuifolia</i> ^d	Herb		+	—	—	49

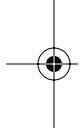
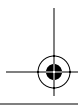
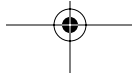
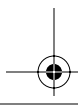




TABLE 21.1 (continued)
Natural Products from Plants that Exhibit Chemopreventive and Therapeutic Activities against Cancer

Compound	Source	Botanical Name	Structure	↓NF-κB	P	T	Ref.
Hematein*	Natural dye from logwood			+	—	—	50,51
Hesperidine	Oranges	<i>Citrus</i> spp.	Hesperitin-7-rutinoside	—	—	—	52
HMP*	Black fruit or galangal	<i>Alpinia</i> spp.	7-(4'-Hydroxy-3'-methoxyphenyl)-phenylheptenone	+	—	—	53
Hypericin*	St. John's wort	<i>Hypericum</i> spp.		+	+	+	54,55
Isothymusin	Basil	<i>Ocimum</i> spp.; <i>Linnophila Geoffrayi</i> ; <i>Bectium grandiflorum</i>	6,7-Dimethoxy-5,8,4'-trihydroxyflavone	—	—	—	56
Isomallotochromanol* and isomallotochromene		<i>Mallotus japonicus</i>		+	—	—	57
Kaempferol	Fruits and vegetables e.g., tomato, onions	<i>Lycopersicon esculentum</i> , <i>Ginkgo biloba</i>	3,5,7,4'-Tetrahydroxyflavone	—	—	—	58
Luteolin*	Fruits and vegetables, tea	<i>Camellia sinensis</i> ; <i>Scutellaria</i> spp.	2-(3,4-Dihydroxyphenyl)-5,7-dihydroxy-chromen-4-one	+	—	—	25,59
Morin	Guava, almond	<i>Psidium guajava</i> ; <i>Prunus dulcis</i>	2',3,4',5,7-Pentahydroxy-flavone	—	+	+	60
Myricetin	Fruits and vegetables		2-(3,4,5-Trihydroxyphenyl)-3,5,7-Trihydroxy-chromen-4-one	—	—	—	61

AU: What does this symbol indicate?



Preventive and Therapeutic Effects of Plant Polyphenols

249

Nasumin	Eggplant	<i>Solanum melongena</i>	Delphinidin-3-(<i>p</i> -coumaroylrutinoside)-5-glucoside	—	—	—	62
Nobiletin	Citrus	<i>Citrus</i> spp.	5,6,7,8,3',4'-Hexamethoxyflavone	—	—	—	28
Nordihydroguaiaretic acid*		<i>Guaiacum officinale</i>		+	—	—	63
<i>Ochna macrocalyx</i> ext.		<i>Ochna macrocalyx</i>		+	—	—	64
Oenotherin B		<i>Oenothera</i> spp.; <i>Eugenia uniflora</i>	Hydrolyzable ellagitannin	—	—	—	65
Panduratin A		<i>Kaempferia pandurata</i>		+	—	—	66
Procyanidins	Tea, cranberries, apple, grape seeds, pear	<i>Camellia sinensis</i> , <i>Vaccinium</i> spp.; <i>Prunus</i> spp.	Condensed tannins	—	—	—	67
Purpurogallin		<i>Quercus</i> sp. Nutgall		—	—	—	68,69
Pycnogenol	Maritime pine bark extract	<i>Pinus maritima</i>	2,3,4,6-Tetrahydroxy-5H-bezocyclohepten-5-one	+	—	—	70,71
Quercetin	Fruits and vegetables	<i>Malus</i> spp.; <i>Lycopersicon esculentum</i>	Bioflavanoid extract	+	—	—	72
Rhein		<i>Daylilies (Hemerocallis</i> spp.); <i>Rheum officinale</i> (<i>dahuang</i>)	2-(3,4-Dihydroxyphenyl)-3,5,7-trihydroxy-chromen-4-one	+	—	—	73
Sangenon C*	Mulberry	<i>Morus</i> spp.	1,8-Dihydroxy-3-carboxyanthraquinone	+	—	—	74
Silymarin ^e	Milk thistle, artichokes	<i>Silybum marianum</i> ; <i>Cynara scolymus</i>	Flavonolignan extract	+	+	+	75,76,77
Saucerneols*, sauchinone, and manassantins*		<i>Saururus</i> spp.		+	—	—	78,79
Tangeretin	citrus fruits	<i>Citrus</i> spp.	5,6,7,8,4'-Pentamethoxyflavone	—	—	—	80

TABLE 21.1 (continued)
Natural Products from Plants that Exhibit Chemopreventive and Therapeutic Activities against Cancer

Compound	Source	Botanical Name	Structure	↓NF-κB	P	T	Ref.
Wedelolactone*		<i>Wedelia</i> spp.; <i>Eclipta alba</i>	1,8,9-Trihydroxy-3-methoxy-6H-benzofuro[3,2-f][1H]-benzopyran-6-one,	+	—	—	81
Yakuchinones* A and B		<i>Alpinia oxyphylla</i>	1-(4'-Hydroxy-3'-methoxyphenyl)-7-phenyl-3-heptanone, 1-(4'-hydroxy-3'-methoxyphenyl)-7-phenylhept-1-en-3-one	+	—	—	82
Terpenes							
Andalusol*		<i>Siderites foetens</i>		+	—	—	83
Anethol [†] and analogs	Broccoli, anise, cloves, cashew	<i>Brassica oleracea</i> ; <i>Illicium verum</i> ; <i>Ocimum</i> spp.; <i>Syzygium aromaticum</i> ; <i>Anacardium occidentale</i> ; <i>Hibiscus sabdariffa</i>	4-Methoxypropenylbenzene	+	+	+	84,85
Artemisinin (qinghaosu)		<i>Artemisia annua</i> L. spp.		+	—	—	86
Avicins* ^g		<i>Acacia victoriae</i>		+	—	—	87
Azadirachtin ^h	Neem tree	<i>Azadirachta indica</i> , <i>A. Jussieu</i>		—	—	—	88
β-carotene	Carrot, citrus fruits, pumpkin	<i>Daucus carota sativus</i> ; <i>Citrus unshiu</i> mar; <i>Curcubita moschata</i>		—	—	—	89,90
β-cryptoxanthin	Fruits	<i>Carica papaya</i> L. <i>Physalis</i>	β,β-Caroten-3-ol	—	—	—	89,91
Bakuchiol (drupanol)*	Bemchi seeds	<i>Psoralea corylifolia</i> (bemchi); <i>Otholobium pubescens</i>	4-(3-ethenyl-3,7-dimethyl-1,6-octadienyl)-phenol	+	—	—	92

AU: What does this symbol indicate?

Preventive and Therapeutic Effects of Plant Polyphenols

Betulinic acid*	Birch tree, almond hulls	<i>Betula</i> spp.; <i>Quisqualis Fructus</i> ; <i>Coussarea paniculata</i> ; <i>Alangium lamarckii</i>	3- β -Hydroxy-lup-20(29)en-28-acid	+	—	—	93
Carnosol*	Rosemary, sage	<i>Rosmarinus officinalis</i> ; <i>Salvia officinalis</i>	2H-9,4a-(epoxymethano)-phenanthren-12-one	+	—	—	94
Celastrol*		<i>Celastrus orbiculatus</i>		+	—	—	95
Costunolide*		<i>Magnolia grandiflora</i> ; <i>Tsoongiodendron odorum</i> ; <i>Saussurea lappa</i>		+	—	—	96
Cucurbitacins ¹	Cucurbitaceae	<i>Cucurbita andreana</i> ; <i>Trichosanthes kirilowii</i> ; <i>Elaeocarpus mastersii</i>	3- β -(β -D-Glucosyloxy)-16-,23- α -epoxycucurbita-5,24-diene-11-one	—	—	—	97,98
Ergolide*		<i>Inula</i> spp.	Dihydrobigelovin	+	—	—	99
Excisanin A*		<i>Isodon (Rabdosia)</i> spp.		+	—	—	100
Foliol*		<i>Sideritis</i> spp.		+	—	—	101
Germacranolides * ¹ and Eudesmanolides		<i>Carpesium divaricatum</i> ; <i>Montanoa hibiscifolia</i>		+	—	—	102,103
Ginkgo biloba ext.		<i>Ginkgo biloba</i>		+	—	—	104
Ginsenoside Rg3*		<i>Panax</i> spp.		+	—	—	105
Glycyrrhizin*	Licorice root	<i>Glycyrrhiza glabra</i> ; <i>Glycyrrhiza uralens</i>		+	—	—	106
Guaianolides*		<i>Viguiera gardneri</i>		+	—	—	107
Helenalin*		<i>Arnicae</i> spp.; <i>Helenium aromaticum</i>		+	—	—	108
Hypoestoxide		<i>Hypoestes rosea</i>		+	—	—	109,110
Kamebacetal A*		<i>Isodon (Rabdosia)</i> spp.		+	—	—	100
Kamebakaurin		<i>Isodon (Rabdosia)</i> spp.		+	—	—	100
Kaurenic acid*		<i>Sideritis</i> spp.	ent-kaur-16-ene-19-oic acid	+	—	—	101
Limonene	Citrus fruits	<i>Citrus</i> spp.	4-Isopropenyl-1-methyl-1-cyclohexane	—	+	+	111

AU: Is this complete?



TABLE 21.1 (continued)
Natural Products from Plants that Exhibit Chemopreventive and Therapeutic Activities against Cancer

Compound	Source	Botanical Name	Structure	↓NF-κB	P	T	Ref.
Linacrol*		<i>Sideritis</i> spp.		+	—	—	101
Lutein	Tomato	<i>Lycopersicon esculentum</i>		—	—	—	112,113
Lycopene	Tomato	<i>Lycopersicon esculentum</i>	ψ,ψ-Carotene	—	—	—	112–114
Oleandrin*		<i>Nerium oleander</i> ; <i>Plumeria obsta</i>		+	—	—	115
Oxoacanthospermoides [§]		<i>Milleria quinqueflora</i>		+	—	—	116
Parthenolide*	Feverfew	<i>Tanacetum parthenium</i> and <i>T. larvatum</i> ; <i>Michelia champaca</i> ; <i>Talauma ovata</i> ; <i>Magnolia grandiflora</i> ; <i>Artemisia myriantha</i>		+	—	—	117,118
Pristimerin*		<i>Hippocratea</i> spp.; <i>Maytenus</i> spp.; <i>Celastrus orbiculatus</i> ; <i>Reissantia buchananii</i> ; <i>Salacia</i> <i>beddomei</i> ; <i>Heisteria pallida</i>		+	—	—	119
Triptolide** (PG 490)		<i>Tripterygium wilfordii</i>		+	—	—	120,121
Ursolic acid*	Basil, rosemary, berries	<i>Rosmarinus officinalis</i> ; <i>Ocimum</i> <i>sanctum</i> ; <i>Aronia melanocarpa</i> ; <i>Oxycoccus quadripetalus</i> ; <i>Origanum</i> <i>majorana</i> ; <i>Diospyros melanoxylon</i> ; <i>Salvia przewalskii</i> Maxim		+	—	—	122
Withanolides	Solanaceae	<i>Withania</i> spp.; <i>Physalis angulata</i> ; <i>Soripichroa origanifolia</i>		—	—	—	123,124

AU: What does this symbol indicate?

AU: Is this correct?

AU: Should this be "Salpichroa"?

Preventive and Therapeutic Effects of Plant Polyphenols

		Alkaloids			
Capsaicinoids* ¹	Pepper, red chilli, paprika fruits	<i>Capsicum</i> spp.	+	+	125-127
Cepharanthine*		<i>Stephania cepharantha</i>	+	-	128
Conophylline*		<i>Tabernaemontana</i> spp.; <i>Ervatamia microphylla</i>	+	-	129
Higenamine	Ranunculaceae	<i>Aconitum japonicum</i> ; <i>Argemone mexicana</i> ; <i>Gnetum parvifolium</i>	-	-	130,131
Mahanimbine	Rutaceae	<i>Murraya koenigii</i> ; <i>Clausena dunniana</i> ; <i>Murraya siamensis</i>	-	+	132,133
Mahanine	Rutaceae	<i>Murraya koenigii</i> ; <i>Micromelum minutum</i>	-	-	132
Morphine ^m and its analogs	Opium poppy	<i>Papaver</i> spp.	+	-	134-136
Murrayanol	Rutaceae	<i>Murraya koenigii</i>	-	-	133
Piperine	Black pepper	<i>Garcinia xanthochymus</i> ; <i>Piper</i> spp.	-	-	115,137
Rocoglamides*		<i>Aglata</i> spp.	+	-	138
Tetrandrine*		<i>Stephania tetrandra</i>	+	-	139
(sinomenine A)		<i>Thionia diversifolia</i> ext.	-	-	140,141
Allicin	Garlic	<i>Allium sativum</i>	-	-	142,143
Lapachone	Ginseng, lapacho tree, trunkwood	<i>Tabebuia</i> spp.	+	+	21

8-Methyl-N-vanillyl-trans-6-noneamide

Benzyl-tetrahydroisoquinoline alkaloid

2-Propene-1-sulfinothioic acid-S-2-propenyl ester

Allylthiosulfinate

Benz[a]phenazine



TABLE 21.1 (continued)
Natural Products from Plants that Exhibit Chemopreventive and Therapeutic Activities against Cancer

Compound	Source	Botanical Name	Structure	↓NF-κB	P	T	Ref.
Rotenone		Benzopyrene <i>Derris</i> spp.		+	—	—	144
CAPE	Honey bee propolis	Caffeic acid phenylethyl ester <i>Apis mellifera capensis</i>		+	—	—	21
Pheophorbide A		Chlorophyll Catabolite <i>Solanum difflorum</i>		+	—	—	145
Sulphoraphane	Brassicaceae, e.g., broccoli, cauliflower	Glucosinolate <i>Brassica oleracea</i>	4-Methylsulphonyl butyl - isothiocyanate	—	—	—	146
Indole-3-carbinol	Brassicaceae, e.g., onions, cabbage	Indoles <i>Allium cepa; Brassica</i> spp.	3-Indolemethanol	—	—	—	147
Aucubin*	Algae	Iridoid glycoside <i>Eucommia</i> spp.; <i>Veronica</i> spp.; <i>Vitex</i> spp.; <i>Globularia</i> spp.		+	—	—	148
Plumbagin		Naptoquinone <i>Plumbago zeylanica</i>	5-Hydroxy-2-methyl-1,4- naphthoquinone	—	—	—	149

AU: What
 does this
 symbol indi-
 cate?



Preventive and Therapeutic Effects of Plant Polyphenols

1'-Acetoxychavicol acetate	Zingiberaceae	Phenyl Propanoid <i>Zingiber officinale</i> ; <i>Languias galanga</i>	—	—	—	28,150
Ethyl gallate	Grapes, tea, red maple	Phenolics <i>Paeonia</i> spp.; <i>Sophora japonica</i> ; <i>Vitis vinifera</i> ; <i>Vitellaria paradoxa</i> ; <i>Camellia sinensis</i>	+	—	—	151
Galic acid*	Fruits, e.g., guava	<i>Psidium guajava</i> ; <i>Erodium glaucophyllum</i> ; <i>Melaleuca quinquenervia</i> <i>Zingiber officinale</i>	—	+	—	152
Gingerol	Ginger	<i>Garcinia</i> spp. <i>Heracleum laciniatum</i> ; <i>Ruta graveolens</i>	—	—	—	153
Morellin Sphondin*	Indica fruit	<i>Garcinia</i> spp. <i>Heracleum laciniatum</i> ; <i>Ruta graveolens</i>	—	—	—	154
Rosemarinic acid	Rosemary, sage	Phenolic Acid <i>Rosmarinus officinalis</i> ; <i>Salvia officinalis</i> <i>Sida acuta</i>	—	—	—	156
Synapic acid		4-Hydroxy-3,5-dimethoxycinnamic acid	—	—	—	157
Syringic acid		4-Hydroxy-3,5-dimethoxybenzoic acid	—	—	—	158,159
Ganoderma lucidum ext.	Reishi	Polysaccharide <i>Ganoderma lucidum</i>	+	+	+	160

TABLE 21.1 (continued)
Natural Products from Plants that Exhibit Chemopreventive and Therapeutic Activities against Cancer

Compound	Source	Botanical Name	Structure	↓NF-κB	P	T	Ref.
Garcinol ^a and its analogs	<i>Garcinia indica</i> fruit	Polyprenylated Benzophenone Derivatives <i>Garcinia</i> spp.		—	—	—	161
Allixin	Garlic	Phytoalexin <i>Allium sativum</i>	3-Hydroxy-5-methoxy-6-methyl-2-pentyl-4H-pyran-4-one	—	—	—	162
Calagualine		Saponin <i>Polypodium</i> spp.		+	—	—	163
Resveratrol ^a and analogs	<i>Japanese knotweed</i> ; berry fruits, E.grapes, cranberries etc.	Stilbene <i>Polygonum cuspidatum</i> ; <i>Veratrum</i> spp.; <i>Vitis</i> spp.; <i>Vaccinium</i> spp.	<i>trans</i> -3,4',5'-Trihydroxystilbene	+	+	+	164–166
Aged garlic ext. α-lipoic acid ^{aP}	Garlic Asparagus, wheat, potatoes	Others <i>Allium sativum</i>		+	—	—	167,168
Apple ext. (juice) Astaxanthin*	Apple juice Microalga, algae	<i>Malus</i> spp. <i>Haematococcus pluvialis</i>	1,2-Dithiolanepentanoic acid 3,3'-Dihydroxy-β,β,carotene-4,4'-dione	+	—	—	169–171
				+	—	—	172
				+	—	—	173

AU: What does this symbol indicate?

Preventive and Therapeutic Effects of Plant Polyphenols

β-Glucan	Barley, soy bean, mushroom	<i>Avena sativa</i> ; <i>Hordeum vulgare</i> ; <i>Agaricus blazei</i>	—	—	174
β-sitosterol	Plants, nuts, lapacha tree, cactus	<i>Glycyne max</i> ; <i>Arachis</i> spp.; <i>Miconia rubiginosa</i> ; <i>Opuntia ficus-indica</i>	—	—	175
Cat's claw	Basil, thyme	<i>Uncaria tomentosa</i>	+	—	176
Cirsilineol		<i>Ocimum sanctum</i> ; <i>Lantana montevidensis</i> Briq.; <i>Thymus vulgaris</i>	—	—	56
Diallylsulfide	Garlic, Chinese leek	<i>Allium sativum</i>	—	—	177
Flavokawains (Kava lactones)	Kava kava	<i>Piper methysticum</i>	—	—	178
Germinated barley			+	—	179
Perisone A	Avocado	<i>Persea americana</i>	—	—	180
Pomegranate wine			—	—	181
S-allylcysteine*		<i>Punica granatum</i>	—	—	182
Stinging nettle ext.		<i>Allium sativum</i>	+	—	183
Trans-Asarone	Carrot	<i>Urtica dioica</i>	—	—	184
Vitamin C*	Fruits and vegetables	<i>Daucus carota</i> L.	—	—	184
Vitamin E	Plant seeds and vegetables		+	—	185,186
			+	—	187,188

Note: T and P refer to therapy and prevention, respectively; the asterisk indicates that the Chemical structure is shown in Figure 21.2 (A to D).

^a Baicalein and its derivatives include baicalin, wogonin, and wogonin, 6-methoxy-baicalein (oroxylin A).

^b Catechins include catechin, epicatechin, epicatechingallate, epigallocatechin, and epigallocatechingallate; theaflavins are polyphenols found in fermented green tea, i.e., black tea.

^c Synthetic compound closely related to a polyphenol isolated from the Indian plant, *Dysoxylum binectariferum*.

^d *Glossogyne tenuifolia* is Chinese medicine Hiang-ju.

AU: What do the blank spaces indicate?



**TABLE 21.1 (continued)
Natural Products from Plants that Exhibit Chemopreventive and Therapeutic Activities against Cancer**

- ^e Silymarin includes silybin, silibinin, silidian, and silychrist.
- ^f Anethol and analogs include eugenol, bis-eugenol, isoeugenol, and anetholdithiolthione.
- ^g Avicins include avicin D and avicin G.
- ^h Azadirachtin analogs include axadirachtin A,B, D, H, I, etc.
- ⁱ Curcubitacins analogs include curcubitacin B, D, E, etc.
- ^j Germacranolides include 2b,5-epoxy-5,10-dihydroxy-6a-angeloyloxy-9b-isobutyloxy-germacran-8a,12-olide.
- ^k Oxoacanthospermolides include methoxymiller-9Z-enolide.
- ^l Capsaicinoids include capsaicin* and analogs, e.g., resimiferatoxin* (daphnetoxin).
- ^m Morphine and its analogs include KT 90 and sanguinarine.
- ⁿ Garcinol and its analogs include isogarcinol.
- ^o Resveratrol and analogs include piceatannol.
- ^p α -Lipoic acid includes dihydrolipoic acid.



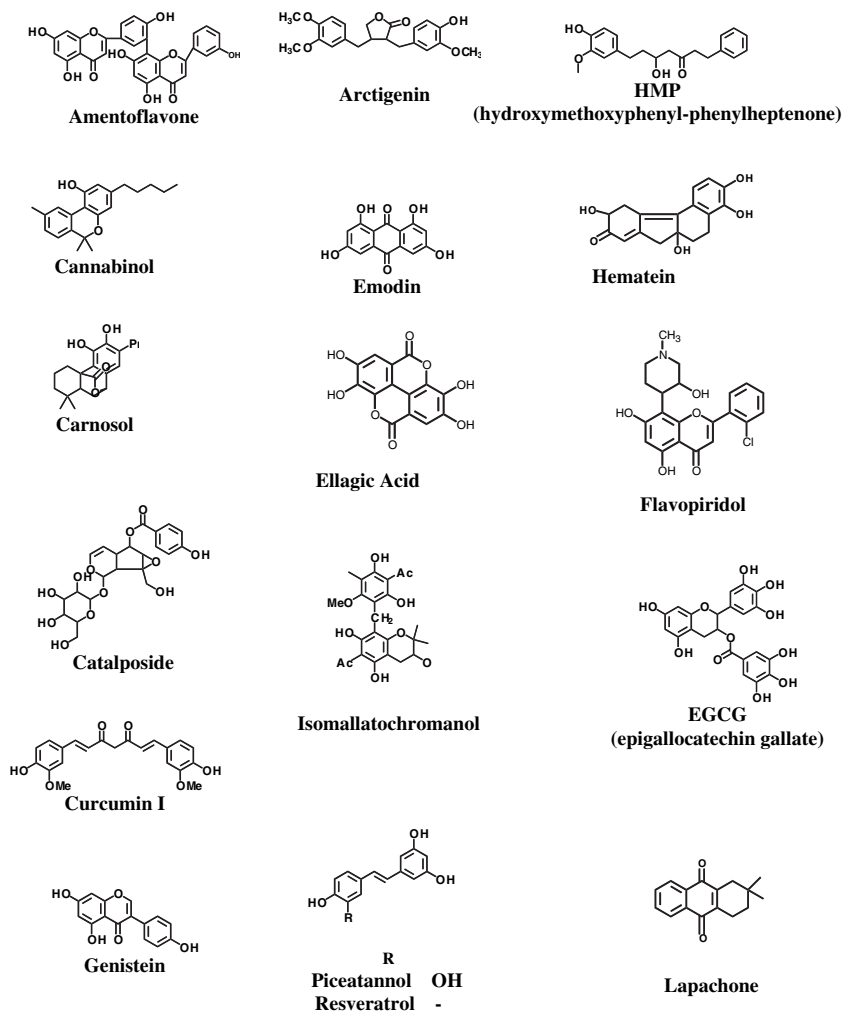
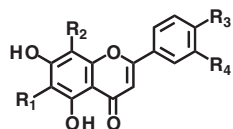
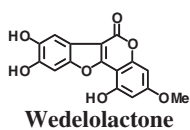
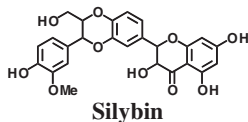
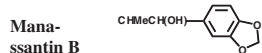
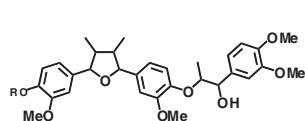
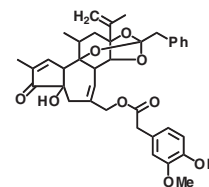
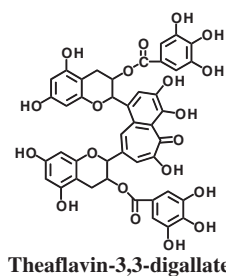
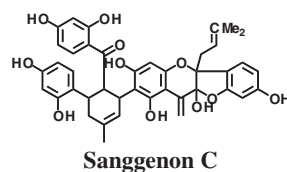
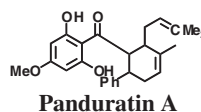
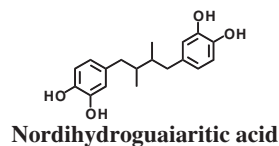


FIGURE 21.2A Structure of some plant-derived polyphenols that block NF- κ B.

that the synergies and additive effects of such treatments, coupled with their mild nature, allow for their safe use. The increase in the popularity and awareness of complementary medicine has resulted in its use for the therapy of a variety of human diseases and by a large percentage of cancer patients. For many such patients, the use of naturally derived plant compounds and plant extracts is an essential part of their treatment. In addition, people are now more aware of the health benefits associated with the use of natural products and plant-derived compounds and have turned to these because of the negative perceptions associated with synthetic compounds. Most of the compounds outlined in Table 21.1 are commonly used in the form of concentrated plant extracts, and combinations



	R ₁	R ₂	R ₃	R ₄
Apigenin	-	-	OH	-
Baicalein	OH	-	-	-
Luteolin	-	-	OH	OH
Oroxylin-A	OCH ₃	-	-	-
Wogonin	-	OCH ₃	-	-



Resiniferatoxin

FIGURE 21.2B Structure of some plant-derived terpenoids that block NF-κB.

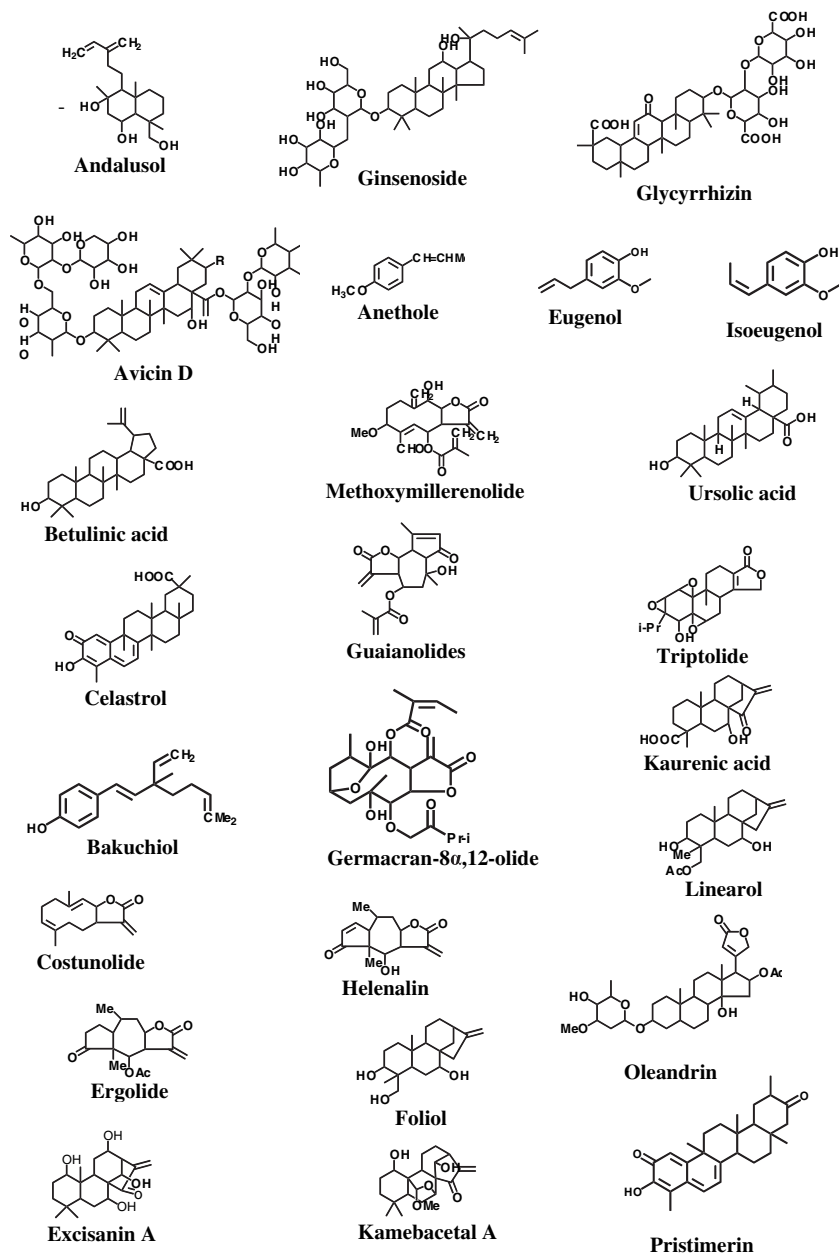


FIGURE 21.2C Structure of some plant-derived alkaloids that block NF-κB.



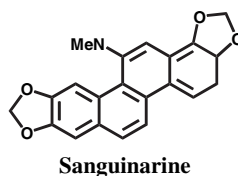
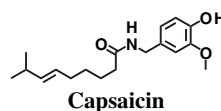
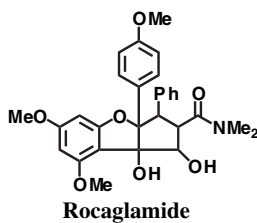
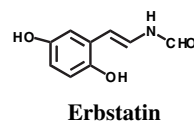
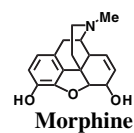
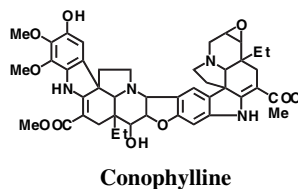
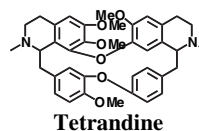
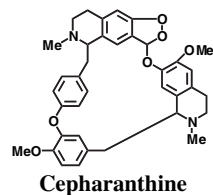
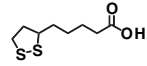


FIGURE 21.2D Structure of some plant-derived miscellaneous compounds that block NF- κ B.

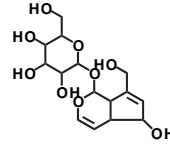
of these extracts can be very complex. However, although humans have been using complex mixtures for much longer than they have been using single isolated compounds or drugs, there is a need for careful standardization of dietary supplements and effective regulatory control to ensure human safety.

ACKNOWLEDGMENTS

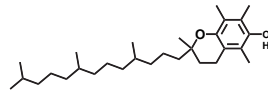
We would like to thank Walter Pagel for a careful review of the manuscript. The contributions of Aggarwal, a Ransom Horne, Jr., Distinguished Professor of Cancer Research, have been supported by the Clayton Foundation for Research,



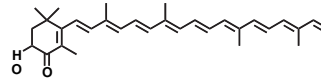
α -lipoic acid



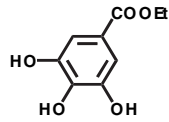
Aucubin



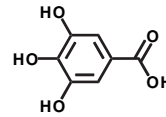
α -tocopherol



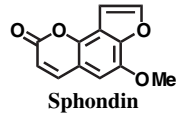
Astaxanthin



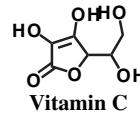
Ethylgallate



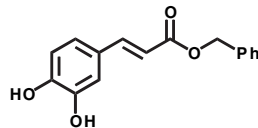
Gallic acid



Sphondin



Vitamin C



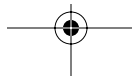
CAPE
(caffeic acid phenylethyl ester)

AU: Pls. provide caption **FIGURE 21.2E** for Fig. 21.2E.

a Department of Defense U.S. Army Breast Cancer Research Program grant (BC010610), a PO1 grant (CA91844) from the National Institutes of Health on lung chemoprevention, and a P50 Head and Neck SPORE grant from the National Institutes of Health.

ABBREVIATIONS

$\text{I}\kappa\text{B}\alpha$: inhibitory subunit of NF- κB
IKK: $\text{I}\kappa\text{B}$ kinase





REFERENCES

AU: Update?

1. Garg A, Aggarwal BB. Nuclear transcription factor- κ B as a target for cancer drug development. *Leukemia* 2002; 16: 1053–1068.
2. Kumar A, Takada Y, Boriek AM, Aggarwal BB. Nuclear factor- κ B: Its role in health and diseases. *J Mol Med* 2004(in press).
3. Shishodia S, Aggarwal BB. Nuclear factor (NF)- κ B regulates the expression of genes involved in transformation, proliferation, invasion, angiogenesis and metastasis of cancer. In *Molecular Targeting and Signal Transduction*. Ed., Rakesh Kumar PD; 2003; Kluwer Publishers: Belgium.
4. Lin A, Karin M. NF- κ B in cancer: a marked target. *Semin Cancer Biol* 2003; 13: 107–114.
5. Orłowski RZ, Baldwin AS, Jr. NF- κ B as a therapeutic target in cancer. *Trends Mol Med* 2002; 8: 385–389.
6. Valen G, Yan ZQ, Hansson GK. Nuclear factor κ -B and the heart. *J Am Coll Cardiol* 2001; 38: 307–314.
7. Jones WK, Brown M, Ren X, He S, McGuinness M. NF- κ B as an integrator of diverse signaling pathways: the heart of myocardial signaling? *Cardiovasc Toxicol* 2003; 3: 229–254.
8. Shoelson SE, Lee J, Yuan M. Inflammation and the IKK beta/I kappa B/NF- κ B axis in obesity- and diet-induced insulin resistance. *Int J Obes Relat Metab Disord* 2003; 27 Suppl. 3: S49–52.
9. Yang L, Cohn L, Zhang DH, Homer R, Ray A, Ray P. Essential role of nuclear factor κ B in the induction of eosinophilia in allergic airway inflammation. *J Exp Med* 1998; 188: 1739–1750.
10. Das J, Chen CH, Yang L, Cohn L, Ray P, Ray A. A critical role for NF- κ B in GATA3 expression and TH2 differentiation in allergic airway inflammation. *Nat Immunol* 2001; 2: 45–50.
11. Gagliardo R, Chanez P, Mathieu M, et al. Persistent activation of nuclear factor- κ B signaling pathway in severe uncontrolled asthma. *Am J Respir Crit Care Med* 2003; 168: 1190–1198.
12. Roshak AK, Callahan JF, Blake SM. Small-molecule inhibitors of NF- κ B for the treatment of inflammatory joint disease. *Curr Opin Pharmacol* 2002; 2: 316–321.
13. van Heel DA, Udalova IA, De Silva AP, et al. Inflammatory bowel disease is associated with a TNF polymorphism that affects an interaction between the OCT1 and NF(- κ)B transcription factors. *Hum Mol Genet* 2002; 11: 1281–1289.
14. Huang CJ, Nazarian R, Lee J, Zhao PM, Espinosa-Jeffrey A, de Vellis J. Tumor necrosis factor modulates transcription of myelin basic protein gene through nuclear factor κ B in a human oligodendrogloma cell line. *Int J Dev Neurosci* 2002; 20: 289–296.
15. Mattson MP, Camandola S. NF-kappaB in neuronal plasticity and neurodegenerative disorders. *J Clin Invest* 2001; 107: 247–254.
16. Kaltschmidt B, Uherek M, Volk B, Baeuerle PA, Kaltschmidt C. Transcription factor NF- κ B is activated in primary neurons by amyloid beta peptides and in neurons surrounding early plaques from patients with Alzheimer disease. *Proc Natl Acad Sci USA* 1997; 94: 2642–2647.
17. Burke JR. Targeting I kappa B kinase for the treatment of inflammatory and other disorders. *Curr Opin Drug Discov Dev* 2003; 6: 720–728.





18. Yamamoto Y, Gaynor RB. Role of the NF- κ B pathway in the pathogenesis of human disease states. *Curr Mol Med* 2001; 1: 287–296.
19. Yamamoto Y, Gaynor RB. Therapeutic potential of inhibition of the NF- κ B pathway in the treatment of inflammation and cancer. *J Clin Invest* 2001; 107: 135–42.
20. Shobana S, Naidu KA. Antioxidant activity of selected Indian spices. *Prostaglandins Leukot Essent Fatty Acids* 2000; 62: 107–110.
21. Natarajan K, Singh S, Burke TR, Jr., Grunberger D, Aggarwal BB. Caffeic acid phenethyl ester is a potent and specific inhibitor of activation of nuclear transcription factor NF- κ B. *Proc Natl Acad Sci USA* 1996; 93: 9090–9095.
22. Banerjee T, Valacchi G, Ziboh VA, van der Vliet A. Inhibition of TNF α -induced cyclooxygenase-2 expression by amentoflavone through suppression of NF- κ B activation in A549 cells. *Mol Cell Biochem* 2002; 238: 105–110.
23. Gupta S, Afaq F, Mukhtar H. Involvement of nuclear factor- κ B, Bax and Bcl-2 in induction of cell cycle arrest and apoptosis by apigenin in human prostate carcinoma cells. *Oncogene* 2002; 21: 3727–3738.
24. Shukla S, Gupta S. Molecular mechanisms for apigenin-induced cell-cycle arrest and apoptosis of hormone refractory human prostate carcinoma DU145 cells. *Mol Carcinog* 2004; 39: 114–126.
25. Choi JS, Choi YJ, Park SH, Kang JS, Kang YH. Flavones mitigate tumor necrosis factor- α -induced adhesion molecule upregulation in cultured human endothelial cells: role of nuclear factor- κ B. *J Nutr* 2004; 134: 1013–1019.
26. Kang BY, Chung SW, Kim SH, Cho D, Kim TS. Involvement of nuclear factor- κ B in the inhibition of interleukin-12 production from mouse macrophages by baicalein, a flavonoid in *Scutellaria baicalensis*. *Planta Med* 2003; 69: 687–691.
27. Cho MK, Park JW, Jang YP, Kim YC, Kim SG. Potent inhibition of lipopolysaccharide-inducible nitric oxide synthase expression by dibenzylbutyrolactone lignans through inhibition of I- κ B α phosphorylation and of p65 nuclear translocation in macrophages. *Int Immunopharmacol* 2002; 2: 105–116.
28. Murakami A, Matsumoto K, Koshimizu K, Ohigashi H. Effects of selected food factors with chemopreventive properties on combined lipopolysaccharide- and interferon- γ -induced I κ B degradation in RAW264.7 macrophages. *Cancer Lett* 2003; 195: 17–25.
29. Krakauer T, Li BQ, Young HA. The flavonoid baicalin inhibits superantigen-induced inflammatory cytokines and chemokines. *FEBS Lett* 2001; 500: 52–55.
30. Kim H, Kim YS, Kim SY, Suk K. The plant flavonoid wogonin suppresses death of activated C6 rat glial cells by inhibiting nitric oxide production. *Neurosci Lett* 2001; 309: 67–71.
31. Chen Y-C, Shen S-C, Hsu F-L. Biological activities of flavonoids. In: *Oriental Foods and Herbs*, ACS Symposium Series, 2003; Vol. 859.
32. Huang C, Huang Y, Li J, et al. Inhibition of benzo(a)pyrene diol-epoxide-induced transactivation of activated protein 1 and nuclear factor κ B by black raspberry extracts. *Cancer Res* 2002; 62: 6857–6863.
33. Atalay M, Gordillo G, Roy S, et al. Anti-angiogenic property of edible berry in a model of hemangioma. *FEBS Lett* 2003; 544: 252–7.
34. Kresty LA, Morse MA, Morgan C, et al. Chemoprevention of esophageal tumorigenesis by dietary administration of lyophilized black raspberries. *Cancer Res* 2001; 61: 6112–6119.



35. Herring AC, Koh WS, Kaminski NE. Inhibition of the cyclic AMP signaling cascade and nuclear factor binding to CRE and kappaB elements by cannabinol, a minimally CNS-active cannabinoid. *Biochem Pharmacol* 1998; 55: 1013–1023.
36. Herring AC, Kaminski NE. Cannabinol-mediated inhibition of nuclear factor- κ B, cAMP response element-binding protein, and interleukin-2 secretion by activated thymocytes. *J Pharmacol Exp Ther* 1999; 291: 1156–1163.
37. Hollister LE. Interactions of marijuana and THC with other drugs: what we don't, but should know, *Marihuana and Medicine*, New York, March 20–21, 1998.
38. An SJ, Pae HO, Oh GS, et al. Inhibition of TNF-alpha, IL-1beta, and IL-6 productions and NF- κ B activation in lipopolysaccharide-activated RAW 264.7 macrophages by catalposide, an iridoid glycoside isolated from *Catalpa ovata* G. Don (Bignoniaceae). *Int Immunopharmacol* 2002; 2: 1173–1181.
39. Mackenzie GG, Carrasquedo F, Delfino JM, Keen CL, Fraga CG, Oteiza PI. Epicatechin, catechin, and dimeric procyanidins inhibit PMA-induced NF- κ B activation at multiple steps in Jurkat T cells. *FASEB J* 2004; 18: 167–169.
40. Pianetti S, Guo S, Kavanagh KT, Sonenshein GE. Green tea polyphenol epigallocatechin-3 gallate inhibits Her-2/neu signaling, proliferation, and transformed phenotype of breast cancer cells. *Cancer Res* 2002; 62: 652–655.
41. Pan MH, Lin-Shiau SY, Ho CT, Lin JH, Lin JK. Suppression of lipopolysaccharide-induced nuclear factor-kappaB activity by theaflavin-3,3'-digallate from black tea and other polyphenols through down-regulation of I κ B kinase activity in macrophages. *Biochem Pharmacol* 2000; 59: 357–367.
42. Ibanez E, Kubatova A, Senorans FJ, Cavero S, Reglero G, Hawthorne SB. Subcritical water extraction of antioxidant compounds from rosemary plants. *J Agric Food Chem* 2003; 51: 375–382.
43. van't Land B, Blijlevens NM, Marteiijn J, et al. Role of curcumin and the inhibition of NF- κ B in the onset of chemotherapy-induced mucosal barrier injury. *Leukemia* 2004; 18: 276–284.
44. Rao CV, Tokumo K, Rigotty J, Zang E, Kelloff G, Reddy BS. Chemoprevention of colon carcinogenesis by dietary administration of piroxicam, alpha-difluoromethylornithine, 16 alpha-fluoro-5-androsten-17-one, and ellagic acid individually and in combination. *Cancer Res* 1991; 51: 4528–4534.
45. Kumar A, Dhawan S, Aggarwal BB. Emodin (3-methyl-1,6,8-trihydroxyanthraquinone) inhibits TNF-induced NF-kappaB activation, I κ B degradation, and expression of cell surface adhesion proteins in human vascular endothelial cells. *Oncogene* 1998; 17: 913–918.
46. Takada Y, Aggarwal BB. Flavopiridol inhibits NF-kappaB activation induced by various carcinogens and inflammatory agents through inhibition of I κ B α kinase and p65 phosphorylation: abrogation of cyclin D1, cyclooxygenase-2, and matrix metalloprotease-9. *J Biol Chem* 2004; 279: 4750–4759.
47. Muraoka K, Shimizu K, Sun X, et al. Flavonoids exert diverse inhibitory effects on the activation of NF-kappaB. *Transplant Proc* 2002; 34: 1335–1340.
48. Gong L, Li Y, Nedeljkovic-Kurepa A, Sarkar FH. Inactivation of NF- κ B by genistein is mediated via Akt signaling pathway in breast cancer cells. *Oncogene* 2003; 22: 4702–4709.
49. Wu MJ, Wang L, Ding HY, Weng CY, Yen JH. *Glossogyne tenuifolia* acts to inhibit inflammatory mediator production in a macrophage cell line by downregulating LPS-induced NF- κ B. *J Biomed Sci* 2004; 11: 186–199.

AU: Not cited
in the text or
table.



50. Choi JH, Jeong TS, Kim DY, et al. Hematein inhibits atherosclerosis by inhibition of reactive oxygen generation and NF- κ B-dependent inflammatory mediators in hyperlipidemic mice. *J Cardiovasc Pharmacol* 2003; 42: 287–295.
51. Hong JJ, Jeong TS, Choi JH, et al. Hematein inhibits tumor necrotic factor- α -induced vascular cell adhesion molecule-1 and NF- κ B-dependent gene expression in human vascular endothelial cells. *Biochem Biophys Res Commun* 2001; 281: 1127–1133.
52. Behar A, Pujade-Lauraine E, Maurel A, et al. The pathophysiological mechanism of fluid retention in advanced cancer patients treated with docetaxel, but not receiving corticosteroid comedication. *Br J Clin Pharmacol* 1997; 43: 653–658.
53. Yadav PN, Liu Z, Rafi MM. A diarylheptanoid from lesser galangal (*Alpinia officinarum*) inhibits proinflammatory mediators via inhibition of mitogen-activated protein kinase, p44/42, and transcription factor nuclear factor- κ B. *J Pharmacol Exp Ther* 2003; 305: 925–931.
54. Bork PM, Bacher S, Schmitz ML, Kaspers U, Heinrich M. Hypericin as a non-antioxidant inhibitor of NF- κ B. *Planta Med* 1999; 65: 297–300.
55. Agostinis P, Vantieghe A, Merlevede W, de Witte PA. Hypericin in cancer treatment: more light on the way. *Int J Biochem Cell Biol* 2002; 34: 221–241.
56. Kelm MA, Nair MG, Strasburg GM, DeWitt DL. Antioxidant and cyclooxygenase inhibitory phenolic compounds from *Ocimum sanctum* Linn. *Phytomedicine* 2000; 7: 7–13.
57. Ishii R, Horie M, Saito K, Arisawa M, Kitanaka S. Inhibition of lipopolysaccharide-induced pro-inflammatory cytokine expression via suppression of nuclear factor- κ B activation by *Mallotus japonicus* phloroglucinol derivatives. *Biochim Biophys Acta* 2003; 1620: 108–118.
58. Lombardi-Boccia G, Lucarini M, Lanzi S, Aguzzi A, Cappelloni M. Nutrients and antioxidant molecules in yellow plums (*Prunus domestica* L.) from conventional and organic productions: a comparative study. *J Agric Food Chem* 2004; 52: 90–94.
59. Kim SH, Shin KJ, Kim D, et al. Luteolin inhibits the nuclear factor- κ B transcriptional activity in Rat-1 fibroblasts. *Biochem Pharmacol* 2003; 66: 955–963.
60. Ono K, Yoshiike Y, Takashima A, Hasegawa K, Naiki H, Yamada M. Potent anti-amyloidogenic and fibril-destabilizing effects of polyphenols in vitro: implications for the prevention and therapeutics of Alzheimer's disease. *J Neurochem* 2003; 87: 172–181.
61. Hollman PC, Katan MB. Health effects and bioavailability of dietary flavonols. *Free Radic Res* 1999; 31 Suppl.: S75–80.
62. Noda Y, Kneyuki T, Igarashi K, Mori A, Packer L. Antioxidant activity of nasunin, an anthocyanin in eggplant peels. *Toxicology* 2000; 148: 119–23.
63. Brennan P, O'Neill LA. Inhibition of nuclear factor κ B by direct modification in whole cells — mechanism of action of nordihydroguaiaritic acid, curcumin and thiol modifiers. *Biochem Pharmacol* 1998; 55: 965–973.
64. Tang S, Bremner P, Kortenkamp A, et al. Biflavonoids with cytotoxic and anti-bacterial activity from *Ochna macrocalyx*. *Planta Med* 2003; 69: 247–253.
65. Aoki K, Maruta H, Uchiumi F, Hatano T, Yoshida T, Tanuma S. A macrocircular ellagitannin, oenothetin B, suppresses mouse mammary tumor gene expression via inhibition of poly(ADP-ribose) glycohydrolase. *Biochem Biophys Res Commun* 1995; 210: 329–337.



66. Yun JM, Kwon H, Hwang JK. In vitro anti-inflammatory activity of panduratin A isolated from *Kaempferia pandurata* in RAW264.7 cells. *Planta Med* 2003; 69: 1102–1108.
67. Llopiz N, Puiggros F, Cespedes E, et al. Antigenotoxic effect of grape seed procyanidin extract in fao cells submitted to oxidative stress. *J Agric Food Chem* 2004; 52: 1083–1087.
68. Wu TW, Zeng LH, Wu J, et al. Molecular structure and antioxidant specificity of purpurogallin in three types of human cardiovascular cells. *Biochem Pharmacol* 1996; 52: 1073–1080.
69. O’Coinceanainn M, Astill C, Baderschneider B. Coordination of aluminium with purpurogallin and theaflavin digallate. *J Inorg Biochem* 2003; 96: 463–468.
70. Peng Q, Wei Z, Lau BH. Pycnogenol inhibits tumor necrosis factor-alpha-induced nuclear factor kappa B activation and adhesion molecule expression in human vascular endothelial cells. *Cell Mol Life Sci* 2000; 57: 834–841.
71. Packer L. Antioxidant and biological activity of a French maritime pine-bark extract. *219th ACS National Meeting*, 2000.
72. Cho SY, Park SJ, Kwon MJ, et al. Quercetin suppresses proinflammatory cytokines production through MAP kinases and NF- κ B pathway in lipopolysaccharide-stimulated macrophage. *Mol Cell Biochem* 2003; 243: 153–160.
73. Martin G, Bogdanowicz P, Domagala F, Ficheux H, Pujol JP. Rhein inhibits interleukin-1 β -induced activation of MEK/ERK pathway and DNA binding of NF- κ B and AP-1 in chondrocytes cultured in hypoxia: a potential mechanism for its disease-modifying effect in osteoarthritis. *Inflammation* 2003; 27: 233–246.
74. Li LC, Shen F, Hou Q, Cheng GF. Inhibitory effect and mechanism of action of sanggenon C on human polymorphonuclear leukocyte adhesion to human synovial cells. *Acta Pharmacol Sin* 2002; 23: 138–142.
75. Dhanalakshmi S, Singh RP, Agarwal C, Agarwal R. Silibinin inhibits constitutive and TNF α -induced activation of NF- κ B and sensitizes human prostate carcinoma DU145 cells to TNF α -induced apoptosis. *Oncogene* 2002; 21: 1759–1767.
76. Manna SK, Mukhopadhyay A, Van NT, Aggarwal BB. Silymarin suppresses TNF-induced activation of NF- κ B, c-Jun N-terminal kinase, and apoptosis. *J Immunol* 1999; 163: 6800–6809.
77. Gallo D, Giacomelli S, Ferlini C, et al. Antitumour activity of the silybin-phosphatidylcholine complex, IdB 1016, against human ovarian cancer. *Eur J Cancer* 2003; 39: 2403–2410.
78. Hwang BY, Lee JH, Nam JB, Hong YS, Lee JJ. Lignans from *Saururus chinensis* inhibiting the transcription factor NF-kappaB. *Phytochemistry* 2003; 64: 765–771.
79. Hwang BY, Lee JH, Jung HS, et al. Sauchinone, a lignan from *Saururus chinensis*, suppresses iNOS expression through the inhibition of transactivation activity of RelA of NF- κ B. *Planta Med* 2003; 69: 1096–1101.
80. Bracke ME, Depypere HT, Boterberg T, et al. Influence of tangeretin on tamoxifen’s therapeutic benefit in mammary cancer. *J Natl Cancer Inst* 1999; 91: 354–359.
81. Kobori M, Yang Z, Gong D, et al. Wedelolactone suppresses LPS-induced caspase-11 expression by directly inhibiting the IKK complex. *Cell Death Differ* 2004; 11: 123–130.
82. Chun KS, Kang JY, Kim OH, Kang H, Surh YJ. Effects of yakuchinone A and yakuchinone B on the phorbol ester-induced expression of COX-2 and iNOS and activation of NF-kappaB in mouse skin. *J Environ Pathol Toxicol Oncol* 2002; 21: 131–139.



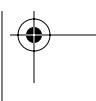
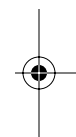
83. de las Heras B, Navarro A, Diaz-Guerra MJ, et al. Inhibition of NOS-2 expression in macrophages through the inactivation of NF- κ B by andalusol. *Br J Pharmacol* 1999; 128: 605–612.
84. Chainy GB, Manna SK, Chaturvedi MM, Aggarwal BB. Anethole blocks both early and late cellular responses transduced by tumor necrosis factor: effect on NF- κ B, AP-1, JNK, MAPKK and apoptosis. *Oncogene* 2000; 19: 2943–2950.
85. Murakami Y, Shoji M, Hanazawa S, Tanaka S, Fujisawa S. Preventive effect of bis-eugenol, a eugenol ortho dimer, on lipopolysaccharide-stimulated nuclear factor κ B activation and inflammatory cytokine expression in macrophages. *Biochem Pharmacol* 2003; 66: 1061–1066.
86. Aldieri E, Atrogene D, Bergandi L, et al. Artemisinin inhibits inducible nitric oxide synthase and nuclear factor NF- κ B activation. *FEBS Lett* 2003; 552: 141–144.
87. Haridas V, Arntzen CJ, Gutterman JU. Avicins, a family of triterpenoid saponins from *Acacia victoriae* (Benthams), inhibit activation of nuclear factor-kappaB by inhibiting both its nuclear localization and ability to bind DNA. *Proc Natl Acad Sci USA* 2001; 98: 11557–11562.
88. Akudugu J, Gade G, Bohm L. Cytotoxicity of azadirachtin A in human glioblastoma cell lines. *Life Sci* 2001; 68: 1153–1160.
89. Adom KK, Sorrells ME, Liu RH. Phytochemical profiles and antioxidant activity of wheat varieties. *J Agric Food Chem* 2003; 51: 7825–7834.
90. Prakash P, Liu C, Hu KQ, Krinsky NI, Russell RM, Wang XD. Beta-carotene and beta-apo-14'-carotenoic acid prevent the reduction of retinoic acid receptor beta in benzo[a]pyrene-treated normal human bronchial epithelial cells. *J Nutr* 2004; 134: 667–673.
91. Uchiyama S, Yamaguchi M. Inhibitory effect of beta-cryptoxanthin on osteoclast-like cell formation in mouse marrow cultures. *Biochem Pharmacol* 2004; 67: 1297–1305.
92. Pae HO, Cho H, Oh GS, et al. Bakuchiol from *Psoralea corylifolia* inhibits the expression of inducible nitric oxide synthase gene via the inactivation of nuclear transcription factor-kappaB in RAW 264.7 macrophages. *Int Immunopharmacol* 2001; 1: 1849–1855.
93. Takada Y, Aggarwal BB. Betulinic acid suppresses carcinogen-induced NF-kappa B activation through inhibition of I kappa B alpha kinase and p65 phosphorylation: abrogation of cyclooxygenase-2 and matrix metalloprotease-9. *J Immunol* 2003; 171: 3278–3286.
94. Lo AH, Liang YC, Lin-Shiau SY, Ho CT, Lin JK. Carnosol, an antioxidant in rosemary, suppresses inducible nitric oxide synthase through down-regulating nuclear factor-kB in mouse macrophages. *Carcinogenesis* 2002; 23: 983–991.
95. Jin HZ, Hwang BY, Kim HS, Lee JH, Kim YH, Lee JJ. Antiinflammatory constituents of *Celastrus orbiculatus* inhibit the NF- κ B activation and NO production. *J Nat Prod* 2002; 65: 89–91.
96. Koo TH, Lee JH, Park YJ, et al. A sesquiterpene lactone, costunolide, from *Magnolia grandiflora* inhibits NF- κ B by targeting I κ B phosphorylation. *Planta Med* 2001; 67: 103–107.
97. Sun IC, Kashiwada Y, Morris-Natschke SL, Lee KH. Plant-derived terpenoids and analogues as anti-HIV agents. *Curr Top Med Chem* 2003; 3: 155–169.
98. Blaskovich MA, Sun J, Cantor A, Turkson J, Jove R, Sebt SM. Discovery of JSI-124 (cucurbitacin I), a selective Janus kinase/signal transducer and activator of transcription 3 signaling pathway inhibitor with potent antitumor activity against human and murine cancer cells in mice. *Cancer Res* 2003; 63: 1270–1279.

99. Whan Han J, Gon Lee B, Kee Kim Y, et al. Ergolide, sesquiterpene lactone from *Inula britannica*, inhibits inducible nitric oxide synthase and cyclo-oxygenase-2 expression in RAW 264.7 macrophages through the inactivation of NF-kappaB. *Br J Pharmacol* 2001; 133: 503–512.
100. Hwang BY, Lee JH, Koo TH, et al. Kaurane diterpenes from *Isodon japonicus* inhibit nitric oxide and prostaglandin E2 production and NF-κB activation in LPS-stimulated macrophage RAW264.7 cells. *Planta Med* 2001; 67: 406–410.
101. Castrillo A, de Las Heras B, Hortelano S, Rodriguez B, Villar A, Bosca L. Inhibition of the nuclear factor kappa B (NF-κB) pathway by tetracyclic kaurene diterpenes in macrophages. Specific effects on NF-κB-inducing kinase activity and on the coordinate activation of ERK and p38 MAPK. *J Biol Chem* 2001; 276: 15854–15860.
102. Kim EJ, Jin HK, Kim YK, et al. Suppression by a sesquiterpene lactone from *Carpesium divaricatum* of inducible nitric oxide synthase by inhibiting nuclear factor-kappaB activation. *Biochem Pharmacol* 2001; 61: 903–910.
103. Muller S, Murillo R, Castro V, Brecht V, Merfort I. Sesquiterpene lactones from *Montanoa hibiscifolia* that inhibit the transcription factor NF-κB. *J Nat Prod* 2004; 67: 622–630.
104. Wei Z, Peng Q, Lau BH, Shah V. Ginkgo biloba inhibits hydrogen peroxide-induced activation of nuclear factor κB in vascular endothelial cells. *Gen Pharmacol* 1999; 33: 369–375.
105. Keum YS, Han SS, Chun KS, et al. Inhibitory effects of the ginsenoside Rg3 on phorbol ester-induced cyclooxygenase-2 expression, NF-κB activation and tumor promotion. *Mutat Res* 2003; 523–524: 75–85.
106. Wang JY, Guo JS, Li H, Liu SL, Zern MA. Inhibitory effect of glycyrrhizin on NF-κB binding activity in CCl4- plus ethanol-induced liver cirrhosis in rats. *Liver* 1998; 18: 180–185.
107. Schorr K, Garcia-Pineros AJ, Siedle B, Merfort I, Da Costa FB. Guaianolides from *Viguiera gardneri* inhibit the transcription factor NF-κB. *Phytochemistry* 2002; 60: 733–740.
108. Lyss G, Schmidt TJ, Merfort I, Pahl HL. Helenalin, an anti-inflammatory sesquiterpene lactone from Arnica, selectively inhibits transcription factor NF-kappaB. *Biol Chem* 1997; 378: 951–961.
109. Ojo-Amaize EA, Kapahi P, Kakkanaiah VN, et al. Hypoestoxide, a novel anti-inflammatory natural diterpene, inhibits the activity of IκB kinase. *Cell Immunol* 2001; 209: 149–157.
110. Ojo-Amaize EA, Nchekwube EJ, Cottam HB, et al. Hypoestoxide, a natural nonmutagenic diterpenoid with antiangiogenic and antitumor activity: possible mechanisms of action. *Cancer Res* 2002; 62: 4007–4014.
111. Gould MN, Moore CJ, Zhang R, Wang B, Kennan WS, Haag JD. Limonene chemoprevention of mammary carcinoma induction following direct in situ transfer of v-Ha-ras. *Cancer Res* 1994; 54: 3540–3543.
112. Chew BP, Park JS. Carotenoid action on the immune response. *J Nutr* 2004; 134: 257S–261S.
113. Khachik F, Beecher GR, Smith JC, Jr. Lutein, lycopene, and their oxidative metabolites in chemoprevention of cancer. *J Cell Biochem Suppl* 1995; 22: 236–246.
114. Liu C, Lian F, Smith DE, Russell RM, Wang XD. Lycopene supplementation inhibits lung squamous metaplasia and induces apoptosis via up-regulating insulin-like growth factor-binding protein 3 in cigarette smoke-exposed ferrets. *Cancer Res* 2003; 63: 3138–3144.

115. Selvendiran K, Senthilnathan P, Magesh V, Sakthisekaran D. Modulatory effect of Piperine on mitochondrial antioxidant system in Benzo(a)pyrene-induced experimental lung carcinogenesis. *Phytomedicine* 2004; 11: 85–89.
116. Castro V, Rungeler P, Murillo R, et al. Study of sesquiterpene lactones from *Milleria quinqueflora* on their anti-inflammatory activity using the transcription factor NF- κ B as molecular target. *Phytochemistry* 2000; 53: 257–263.
117. Hehner SP, Heinrich M, Bork PM, et al. Sesquiterpene lactones specifically inhibit activation of NF-kappa B by preventing the degradation of I kappa B-alpha and I kappa B-beta. *J Biol Chem* 1998; 273: 1288–1297.
118. Kwok BH, Koh B, Ndubuisi MI, Elofsson M, Crews CM. The anti-inflammatory natural product parthenolide from the medicinal herb Feverfew directly binds to and inhibits I κ B kinase. *Chem Biol* 2001; 8: 759–766.
119. Dirsch VM, Kiemer AK, Wagner H, Vollmar AM. The triterpenoid quinonemethide pristimerin inhibits induction of inducible nitric oxide synthase in murine macrophages. *Eur J Pharmacol* 1997; 336: 211–217.
120. Sylvester J, Liacini A, Li WQ, Dehnade F, Zafarullah M. *Tripterygium wilfordii* Hook F extract suppresses proinflammatory cytokine-induced expression of matrix metalloproteinase genes in articular chondrocytes by inhibiting activating protein-1 and nuclear factor- κ B activities. *Mol Pharmacol* 2001; 59: 1196–1205.
121. Qiu D, Kao PN. Immunosuppressive and anti-inflammatory mechanisms of triptolide, the principal active diterpenoid from the Chinese medicinal herb *Tripterygium wilfordii* Hook. f. *Drugs in R&D* 2003; 4: 1–18.
122. Shishodia S, Majumdar S, Banerjee S, Aggarwal BB. Ursolic acid inhibits nuclear factor- κ B activation induced by carcinogenic agents through suppression of I κ B α kinase and p65 phosphorylation: correlation with down-regulation of cyclooxygenase 2, matrix metalloproteinase 9, and cyclin D1. *Cancer Res* 2003; 63: 4375–4383.
123. Park EJ, Pezzuto JM. Botanicals in cancer chemoprevention. *Cancer Metastasis Rev* 2002; 21: 231–255.
124. Misico RI, Song LL, Veleiro AS, et al. Induction of quinone reductase by withanolides. *J Nat Prod* 2002; 65: 677–680.
125. Sancho R, Lucena C, Macho A, et al. Immunosuppressive activity of capsaicinoids: capsiate derived from sweet peppers inhibits NF-kappaB activation and is a potent antiinflammatory compound in vivo. *Eur J Immunol* 2002; 32: 1753–1763.
126. Chen CW, Lee ST, Wu WT, Fu WM, Ho FM, Lin WW. Signal transduction for inhibition of inducible nitric oxide synthase and cyclooxygenase-2 induction by capsaicin and related analogs in macrophages. *Br J Pharmacol* 2003; 140: 1077–1087.
127. Singh S, Natarajan K, Aggarwal BB. Capsaicin (8-methyl-N-vanillyl-6-nonanamide) is a potent inhibitor of nuclear transcription factor- κ B activation by diverse agents. *J Immunol* 1996; 157: 4412–4420.
128. Okamoto M, Ono M, Baba M. Suppression of cytokine production and neural cell death by the anti-inflammatory alkaloid cepharanthine: a potential agent against HIV-1 encephalopathy. *Biochem Pharmacol* 2001; 62: 747–753.
129. Gohda J, Inoue J, Umezawa K. Down-regulation of TNF- α receptors by conophylline in human T-cell leukemia cells. *Int J Oncol* 2003; 23: 1373–1379.
130. Chang YC, Chang FR, Khalil AT, Hsieh PW, Wu YC. Cytotoxic benzophenanthridine and benzylisoquinoline alkaloids from *Argemone mexicana*. *Z Naturforsch [C]* 2003; 58: 521–526.



131. Kang YJ, Lee YS, Lee GW, et al. Inhibition of activation of nuclear factor κ B is responsible for inhibition of inducible nitric oxide synthase expression by higenamine, an active component of aconite root. *J Pharmacol Exp Ther* 1999; 291: 314–320.
132. Tachibana Y, Kikuzaki H, Lajis NH, Nakatani N. Antioxidative activity of carbazoles from *Murraya koenigii* leaves. *J Agric Food Chem* 2001; 49: 5589–5594.
133. Ramsewak RS, Nair MG, Strasburg GM, DeWitt DL, Nitiss JL. Biologically active carbazole alkaloids from *Murraya koenigii*. *J Agric Food Chem* 1999; 47: 444–447.
134. Sueoka E, Sueoka N, Kai Y, et al. Anticancer activity of morphine and its synthetic derivative, KT-90, mediated through apoptosis and inhibition of NF- κ B activation. *Biochem Biophys Res Commun* 1998; 252: 566–570.
135. Rajagopal A, Vassilopoulou-Sellin R, Palmer JL, Kaur G, Bruera E. Hypogonadism and sexual dysfunction in male cancer survivors receiving chronic opioid therapy. *J Pain Symptom Manage* 2003; 26: 1055–1061.
136. Chaturvedi MM, Kumar A, Darnay BG, Chaing GB, Agarwal S, Aggarwal BB. Sanguinarine (pseudochelethrine) is a potent inhibitor of NF- κ B activation, I κ B α phosphorylation, and degradation. *J Biol Chem* 1997; 272: 30129–30134.
137. Rauscher FM, Sanders RA, Watkins JB, 3rd. Effects of piperine on antioxidant pathways in tissues from normal and streptozotocin-induced diabetic rats. *J Biochem Mol Toxicol* 2000; 14: 329–334.
138. Baumann B, Bohnstengel F, Siegmund D, et al. Rocaglamide derivatives are potent inhibitors of NF- κ B activation in T-cells. *J Biol Chem* 2002; 277: 44791–44800.
139. Ye J, Ding M, Zhang X, Rojanasakul Y, Shi X. On the role of hydroxyl radical and the effect of tetrandrine on nuclear factor — κ B activation by phorbol 12-myristate 13-acetate. *Ann Clin Lab Sci* 2000; 30: 65–71.
140. Bork PM, Schmitz ML, Kuhnt M, Escher C, Heinrich M. Sesquiterpene lactone containing Mexican Indian medicinal plants and pure sesquiterpene lactones as potent inhibitors of transcription factor NF- κ B. *FEBS Lett* 1997; 402: 85–90.
141. Bork PM, Schmitz ML, Weimann C, Kist M, Heinrich M. Nahua Indian medicinal plants (Mexico). Inhibitory activity on NF- κ B antibacterial effects. *Phytochemistry* 1996; 3: 263–269.
142. Hirsch K, Danilenko M, Giat J, et al. Effect of purified allicin, the major ingredient of freshly crushed garlic, on cancer cell proliferation. *Nutr Cancer* 2000; 38: 245–254.
143. Oommen S, Anto RJ, Srinivas G, Karunakaran D. Allicin (from garlic) induces caspase-mediated apoptosis in cancer cells. *Eur J Pharmacol* 2004; 485: 97–103.
144. Suzuki YJ, Mizuno M, Packer L. Signal transduction for nuclear factor- κ B activation. Proposed location of antioxidant-inhibitable step. *J Immunol* 1994; 153: 5008–5015.
145. Heinrich M, Bork PM, Schmitz ML, Rimpler H, Frei B, Sticher O. Pheophorbide A from *Solanum diflorum* interferes with NF- κ B activation. *Planta Med* 2001; 67: 156–157.
146. van Lieshout EM, Posner GH, Woodard BT, Peters WH. Effects of the sulforaphane analog compound 30, indole-3-carbinol, D-limonene or relafen on glutathione S-transferases and glutathione peroxidase of the rat digestive tract. *Biochim Biophys Acta* 1998; 1379: 325–336.
147. Chinni SR, Li Y, Upadhyay S, Koppolu PK, Sarkar FH. Indole-3-carbinol (I3C) induced cell growth inhibition, G1 cell cycle arrest and apoptosis in prostate cancer cells. *Oncogene* 2001; 20: 2927–2936.





148. Jeong HJ, Koo HN, Na HJ, et al. Inhibition of TNF- α and IL-6 production by Aucubin through blockade of NF- κ B activation RBL-2H3 mast cells. *Cytokine* 2002; 18: 252–259.
149. Srinivas G, Annab LA, Gopinath G, Banerji A, Srinivas P. Antisense blocking of BRCA1 enhances sensitivity to plumbagin but not tamoxifen in BG-1 ovarian cancer cells. *Mol Carcinog* 2004; 39: 15–25.
150. Nakamura Y, Murakami A, Ohto Y, Torikai K, Tanaka T, Ohigashi H. Suppression of tumor promoter-induced oxidative stress and inflammatory responses in mouse skin by a superoxide generation inhibitor 1'-acetoxychavicol acetate. *Cancer Res* 1998; 58:4832-9.
151. Murase T, Kume N, Hase T, et al. Gallates inhibit cytokine-induced nuclear translocation of NF- κ B and expression of leukocyte adhesion molecules in vascular endothelial cells. *Arterioscler Thromb Vasc Biol* 1999; 19: 1412–1420.
152. Milner JA. Mechanisms by which garlic and allyl sulfur compounds suppress carcinogen bioactivation. Garlic and carcinogenesis. *Adv Exp Med Biol* 2001; 492: 69–81.
153. Bode AM, Ma WY, Surh YJ, Dong Z. Inhibition of epidermal growth factor-induced cell transformation and activator protein 1 activation by [6]-gingerol. *Cancer Res* 2001; 61: 850–853.
154. Sani BP, Rao PL. Antibiotic principles of *Garcinia morella*. VII. Antiprotozoal activity of morellin, neomorellin & other insoluble neutral phenols of the seed coat of *Garcinia morella*. *Indian J Exp Biol* 1966; 4: 27–28.
155. Yang LL, Liang YC, Chang CW, et al. Effects of sphondin, isolated from *Heraclium laciniatum*, on IL-1 β -induced cyclooxygenase-2 expression in human pulmonary epithelial cells. *Life Sci* 2002; 72: 199–213.
156. Kohda H, Takeda O, Tanaka S, et al. Isolation of inhibitors of adenylate cyclase from dan-shen, the root of *Salvia miltiorrhiza*. *Chem Pharm Bull (Tokyo)* 1989; 37: 1287–90.
157. Bylka W, Matlawska I. Flavonoids and free phenolic acids from *Phytolacca americana* L. leaves. *Acta Pol Pharm* 2001; 58: 69–72.
158. Liu Y, Fang J, Lei T, Wang W, Lin A. Anti-endotoxic effects of syringic acid of *Radix isatidis*. *J Huazhong Univ Sci Technol Med Sci* 2003; 23: 206–208.
159. Kampa M, Alexaki VI, Notas G, et al. Antiproliferative and apoptotic effects of selective phenolic acids on T47D human breast cancer cells: potential mechanisms of action. *Breast Cancer Res* 2004; 6: R63–R74.
160. Sliva D, Labarrere C, Slivova V, Sedlak M, Lloyd FP, Jr., Ho NW. *Ganoderma lucidum* suppresses motility of highly invasive breast and prostate cancer cells. *Biochem Biophys Res Commun* 2002; 298: 603–612.
161. Tanaka T, Kohno H, Shimada R, et al. Prevention of colonic aberrant crypt foci by dietary feeding of garcinol in male F344 rats. *Carcinogenesis* 2000; 21: 1183–1189.
162. Borek C. Antioxidant health effects of aged garlic extract. *J Nutr* 2001; 131: 1010S–1015S.
163. Manna SK, Bueso-Ramos C, Alvarado F, Aggarwal BB. Calagualine inhibits nuclear transcription factors- κ B activated by various inflammatory and tumor promoting agents. *Cancer Lett* 2003; 190: 171–182.
164. Manna SK, Mukhopadhyay A, Aggarwal BB. Resveratrol suppresses TNF-induced activation of nuclear transcription factors NF- κ B, activator protein-1, and apoptosis: potential role of reactive oxygen intermediates and lipid peroxidation. *J Immunol* 2000; 164: 6509–6519.





165. Holmes-McNary M, Baldwin AS, Jr. Chemopreventive properties of trans-resveratrol are associated with inhibition of activation of the I κ B kinase. *Cancer Res* 2000; 60: 3477–3483.
166. Ashikawa K, Majumdar S, Banerjee S, Bharti AC, Shishodia S, Aggarwal BB. Piceatannol inhibits TNF-induced NF- κ B activation and NF- κ B-mediated gene expression through suppression of I κ B α kinase and p65 phosphorylation. *J Immunol* 2002; 169: 6490–6497.
167. Ide N, Lau BH. Garlic compounds minimize intracellular oxidative stress and inhibit nuclear factor- κ B activation. *J Nutr* 2001; 131: 1020S–1026S.
168. Keiss HP, Dirsch VM, Hartung T, et al. Garlic (*Allium sativum* L.) modulates cytokine expression in lipopolysaccharide-activated human blood thereby inhibiting NF- κ B activity. *J Nutr* 2003; 133: 2171–2175.
169. Horakova O, Kalamar J, Sopinska M, Horak F. Presence of α -lipoic acid in natural raw materials. *Cesko-Slovenska Farmacie* 1964; 13: 107–110.
170. Sen CK, Tirosch O, Roy S, Kobayashi MS, Packer L. A positively charged alpha-lipoic acid analogue with increased cellular uptake and more potent immunomodulatory activity. *Biochem Biophys Res Commun* 1998; 247: 223–228.
171. Suzuki YJ, Aggarwal BB, Packer L. Alpha-lipoic acid is a potent inhibitor of NF-kappa B activation in human T cells. *Biochem Biophys Res Commun* 1992; 189: 1709–1715.
172. Shi D, Jiang BH. Antioxidant properties of apple juice and its protection against Cr(VI)-induced cellular injury. *J Environ Pathol Toxicol Oncol* 2002; 21: 233–242.
173. Lee SJ, Bai SK, Lee KS, et al. Astaxanthin inhibits nitric oxide production and inflammatory gene expression by suppressing I(kappa)B kinase-dependent NF-kappaB activation. *Mol Cells* 2003; 16: 97–105.
174. Bobek P, Galbavy S. Effect of pleuran (beta-glucan from *Pleurotus ostreatus*) on the antioxidant status of the organism and on dimethylhydrazine-induced precancerous lesions in rat colon. *Br J Biomed Sci* 2001; 58: 164–168.
175. Nigro ND, Bull AW, Wilson PS, Soullier BK, Alousi MA. Combined inhibitors of carcinogenesis: effect on azoxymethane-induced intestinal cancer in rats. *J Natl Cancer Inst* 1982; 69: 103–107.
176. Aguilar JL, Rojas P, Marcelo A, et al. Anti-inflammatory activity of two different extracts of *Uncaria tomentosa* (Rubiaceae). *J Ethnopharmacol* 2002; 81: 271–276.
177. Wargovich MJ, Woods C, Eng VW, Stephens LC, Gray K. Chemoprevention of N-nitrosomethylbenzylamine-induced esophageal cancer in rats by the naturally occurring thioether, diallyl sulfide. *Cancer Res* 1988; 48: 6872–6875.
178. Hashimoto T, Suganuma M, Fujiki H, Yamada M, Kohno T, Asakawa Y. Isolation and synthesis of TNF-alpha release inhibitors from Fijian kava (*Piper methysticum*). *Phytomedicine* 2003; 10: 309–317.
179. Kanauchi O, Serizawa I, Araki Y, et al. Germinated barley foodstuff, a prebiotic product, ameliorates inflammation of colitis through modulation of the enteric environment. *J Gastroenterol* 2003; 38: 134–141.
180. Kim OK, Murakami A, Takahashi D, et al. An avocado constituent, persenone A, suppresses expression of inducible forms of nitric oxide synthase and cyclooxygenase in macrophages, and hydrogen peroxide generation in mouse skin. *Biosci Biotechnol Biochem* 2000; 64: 2504–2507.
181. Schubert SY, Neeman I, Resnick N. A novel mechanism for the inhibition of NF-kappaB activation in vascular endothelial cells by natural antioxidants. *Faseb J* 2002; 16:1931–1933.



Preventive and Therapeutic Effects of Plant Polyphenols

275

182. Geng Z, Rong Y, Lau BH. S-allyl cysteine inhibits activation of nuclear factor κ B in human T cells. *Free Radic Biol Med* 1997; 23: 345–350.
183. Riehemann K, Behnke B, Schulze-Osthoff K. Plant extracts from stinging nettle (*Urtica dioica*), an antirheumatic remedy, inhibit the proinflammatory transcription factor NF- κ B. *FEBS Lett* 1999; 442: 89–94.
184. Momin RA, De Witt DL, Nair MG. Inhibition of cyclooxygenase (COX) enzymes by compounds from *Daucus carota* L. Seeds. *Phytother Res* 2003; 17: 976–979.
185. Carcamo JM, Pedraza A, Borquez-Ojeda O, Golde DW. Vitamin C suppresses TNF alpha-induced NF kappa B activation by inhibiting I kappa B alpha phosphorylation. *Biochemistry* 2002; 41: 12995–13002.
186. Bowie AG, O'Neill LA. Vitamin C inhibits NF-kappa B activation by TNF via the activation of p38 mitogen-activated protein kinase. *J Immunol* 2000; 165: 7180–7188.
187. Siler U, Barella L, Spitzer V, et al. Lycopene and Vitamin E interfere with autocrine/paracrine loops in the Dunning prostate cancer model. *Faseb J* 2004.
188. Egger T, Schuligoi R, Wintersperger A, Amann R, Malle E, Sattler W. Vitamin E (alpha-tocopherol) attenuates cyclo-oxygenase 2 transcription and synthesis in immortalized murine BV-2 microglia. *Biochem J* 2003; 370: 459–467.

