

Nuclear Factor- κ B As Target for Chemoprevention

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ABBREVIATIONS

NF- κ B, nuclear factor kappa B; I κ B, inhibitor of NF- κ B; CAPE, caffeic acid phenethyl ester; PBIT; S,S'-1,4-Phenylene-bis (1,2-ethanediyl) bis-isothiourea; PDTC, pyrrolidine dithiocarbamate.

ABSTRACT

The process of tumorigenesis requires cellular transformation, hyperproliferation, invasion, angiogenesis, and metastasis. Several genes that mediate these processes are regulated by the transcription factor NF- κ B. The latter is activated by

1 various carcinogens, inflammatory agents, and tumor promo-
2 ters. The NF- κ B, a transcription factor, is present normally in
3 the cytoplasm as an inactive heterotrimer consisting of p50,
4 p65, and I κ B α subunits. When activated, NF- κ B translocates
5 to the nucleus as a p50-p65 heterodimer. This factor regulates the
6 expression of various genes that control apoptosis, viral replica-
7 tion, tumorigenesis, various autoimmune diseases, and inflam-
8 mation. The NF- κ B has been linked to the development of
9 carcinogenesis for several reasons. First, various carcinogens
10 and tumor promoters have been shown to activate NF- κ B.
11 Second, activation of NF- κ B has been shown to block apopto-
12 sis and promote proliferation. Third, the tumor microenviron-
13 ment can induce NF- κ B activation. Fourth, constitutive
14 expression of NF- κ B is frequently found in tumor cells. Fifth,
15 NF- κ B activation induces resistance to chemotherapeutic
16 agents. Sixth, several genes involved in tumor initiation, pro-
17 motion, and metastasis are regulated by NF- κ B. Seventh,
18 various chemopreventive agents have been found to downre-
19 gulate the NF- κ B activation. All these observations suggest
20 that NF- κ B could mediate tumorigenesis and thus can be
21 used as a target for chemoprevention and for the treatment
22 of cancer. Agents, which suppress NF- κ B activation, can sup-
23 press the expression of genes involved in carcinogenesis and
24 tumorigenesis in vivo.

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27 I. CARCINOGENESIS/TUMORIGENESIS

28
29 The process of tumorigenesis is a process that requires cellu-
30 lar transformation, hyperproliferation, invasion, angiogen-
31 esis, and metastasis. This process is activated by various
32 carcinogens (such as cigarette smoke), inflammatory agents
33 (such as TNF and H₂O₂), and tumor promoters (such as phor-
34 bol ester and okadaic acid) (1). Although initially identified as
35 an anticancer agent (2), TNF has now been shown to be
36 involved in cellular transformation (3), tumor promotion (4),
37 and induction of metastasis (5–7). In agreement with these
38 observations, mice deficient in TNF have been shown to be
39 resistant to skin carcinogenesis (8). For several tumors,

1 TNF has been shown to be a growth factor (9,10). Like phorbol
2 ester, TNF mediates these effects in part through activation of
3 a protein kinase C pathway (11). Similar to TNF, other inflam-
4 matory cytokines have also been implicated in tumorigenesis
5 (12,13). Thus, agents that can suppress the expression of
6 TNF and other inflammatory agents have chemopreventive
7 potential (14,15). Most carcinogens, inflammatory agents,
8 and tumor promoters including cigarette smoke, phorbol ester,
9 okadaic acid, H₂O₂, and TNF, have been shown to activate the
10 transcription factor NF-κB.

11 12 13 **II. CIGARETTE SMOKE AND CANCER**

14
15 Cigarette smoke(CS) is a major cause of cancers of the lung, AQ4
16 larynx, oral cavity and pharynx, esophagus, pancreas, kidney,
17 and bladder (16). Worldwide, one in seven or 15% (1.1 million
18 new cases per year) of all cancer cases are attributable to CS,
19 25% in men and 4% in women. Recent estimates indicate that
20 CS causes approximately 80–90% of lung cancer in the United
21 States (17). Smoking during pregnancy and passive exposure
22 to CS may increase the risk of cancer for children and adults
23 (18–20). These estimates do not include the disease resulting
24 from smokeless tobacco (taken orally or as snuff), which is a
25 substantial cause of cancer mortality, particularly on the
26 Indian subcontinent (21).

27 Tobacco smoke is a complex mixture containing at least
28 40 different carcinogens, which mediate tumor initiation and
29 promotion. These carcinogens include nitrosamine, polycyclic
30 aromatic hydrocarbons (PAH), aromatic amines, unsaturated
31 aldehydes (e.g., crotonaldehyde), and some phenolic com-
32 pounds (acrolein). The most potent carcinogenic agent
33 contained in CS is the nitrosamine 4-(methylnitrosoamino) -
34 l-(3-pyridyl) -l-butanone (NNK); formed by nitrosation of nico-
35 tine, it is thought to be an important etiological factor in
36 tobacco–smoke related human cancers (22). The NNK is a
37 site-specific carcinogen in that, irrespective of the route of
38 administration, NNK has remarkable specificity for the lung
39 (23). Because side-stream smoke often contains higher

1 amounts of NNK than mainstream smoke, passive exposure to
2 CS has been suggested to be quite harmful (22). An enzyme 11
3 ??-hydroxysteroid dehydrogenase 1(11??-HSD1), which is AQ5
4 involved in metabolism of endogenous steroids, is also respon-
5 sible for the metabolism of NNK. Thus inhibition of 11??-
6 HSD1 can increase the circulating levels of NNK by impairing
7 its metabolism. Ethanol has been shown to be a potent in-
8 hibitor of 11??-HSD (24) and thus may increase the risk of
9 lung cancer for active or passive smokers. An alcohol consump-
10 tion and cigarette smoking have also been shown to increases
11 the frequency of p53, a tumor suppressor gene, mutation in
12 lung cancer (25).

13 Cigarette smoke has been shown to induce aryl hydrocar-
14 bon hydroxylase (AHH) activity, an activator of respiratory
15 tract carcinogens of the PAH (e.g., benzo[a] pyrene) group
16 (26), in human pulmonary macrophages (27) and in patients
17 with smoking-associated malignant cancers (28). It has been
18 postulated that individuals with high activity of oxidative
19 enzymes (cytochrome P-450 enzymes) or a low activity of
20 detoxifying enzymes (e.g., glutathione s-transferase and epox-
21 ide hydroxylase) may be at increased risk for cancer caused by
22 CS (29). Low intake of dietary constituents with antioxidant AQ6
23 properties such as carotene, vitamin C, and vitamin E further
24 increases the cancer risk in smokers (30).

25 Lung tumors from nonsmokers exhibit elevated NAD(P)
26 H:(quinone-acceptor) oxidoreductase (QAO) activity compared
27 to normal tissue, but tumors from smokers show increases in
28 tumor QAO (31). This could influence the response of these
29 tumors to quinone drugs (commonly used to treat cancer) or
30 toxic agents that are metabolized by QAO. Quinone anticancer
31 drugs are activated to alkylating species by reduction to
32 hydroquinone. Metabolism by QAO is responsible for the
33 formation of alkylating species from doxorubicin (32) and
34 other cytotoxic drugs (33).

35 Another possible mechanism by which CS can cause can-
36 cer involves the effects of PAH on the p53 gene. For instance,
37 exposure of cells to benzo(a)pyrene adducts can induce the
38 same mutation in p53 as is found in 60% of all lung cancers
39 (34). Also exposure of cells to PAH and its metabolites results

1 in a rapid accumulation of the p53 gene product (35,36)
2 through activation of a transcription factor, NF-κB (37).
3
4

5 III. EFFECT OF CIGARETTE SMOKE 6 ON PULMONARY INFLAMMATION 7

8 Experimental epidemiological and clinical evidence indicates
9 that CS is a primary risk factor for chronic obstructive
10 pulmonary disease (COPD), which includes chronic bronchitis
11 and emphysema. These two conditions result from obstruction
12 of airflow and usually coexist. An increased proteolytic acti-
13 vity in the lung due to an imbalance between proteases,
14 especially elastase and α₁-1 protease inhibitor (1PI, an antie-
15 lastase), has been suggested as a primary cause for COPD
16 caused by CS. This occurs for three reasons. First, CS causes
17 the generation of chemotactic factors (such as chemokines)
18 (38), which recruit inflammatory cells (such as neutrophils
19 and macrophages) to the lung, and these cells release pro-
20 teolytic enzymes. Second, free radicals present in CS can
21 either inactivate α₁PI by oxidation of an active site methio-
22 nyl residue present in the protein sequence or damage macro-
23 molecules to make them more susceptible to proteolysis.
24 Third, components in CS can suppress elastin synthesis by
25 inhibiting the cross-linking enzyme lysyl oxidase. Thus
26 neutrophil recruitment, inactivation of protease inhibitors,
27 and depressed tissue repair are considered responsible for
28 the pathogenesis of CS-induced emphysema, although, only
29 one in six smokers develop extensive COPD.

30 The inhalation of CS also results in inflammation of the
31 pulmonary epithelia. Reactive oxygen intermediates (ROIs)
32 are some of the most important effector molecules of acute
33 inflammation. The inflammatory cell response to CS has been
34 studied extensively either in cells harvested by bronchoalveo-
35 lar lavage from cigarette smokers or smoke-exposed animals
36 or in macrophages exposed to CS in vitro. Alveolar macro-
37 phages lavaged from smokers have increased oxidative meta-
38 bolism compared to those in nonsmokers, and this leads to
39 increased apoptosis of fibroblasts, which could be prevented

1 by oxidant scavenging agents. Thus oxidants generated by
2 alveolar macrophages from smokers may facilitate tissue
3 destruction (39).
4

5 **IV. OXIDATIVE DAMAGE BY CIGARETTE** 6 **SMOKE** 7

8 Cigarette smoke has been implicated as major risk factor in
9 COPD such as chronic bronchitis and emphysema, in chemical
10 carcinogenesis, and in atherosclerotic arterial diseases. The
11 mechanisms of the adverse biological effects of CS appear, in
12 part, to include oxidative damage to essential biological consti-
13 tuents. The CS increases the number of phagocytes in the blood
14 and lungs (40), decreases plasma levels of high-density lipopro-
15 teins (HDL) (41), and induces lipid peroxidation of LDL (42).
16 Several plasma proteins have been shown to undergo modifica-
17 tion by exposure to CS (43,44). In CS-bubbled buffers, H₂O₂
18 and hydroxyl radical were generated from aqueous extracts
19 of tar (45,46). A superoxide radical was an intermediate in
20 these reactions. Superoxide formed from CS impairs active
21 oxygen generation from neutrophils.
22

23 **V. COMPOSITION OF CIGARETTE SMOKE** 24

25 Cigarette smoke is a complex mixture consisting of tarry
26 particles of respirable size suspended in a mixture of organic
27 and inorganic gases and containing more than 4000 chemical
28 compounds. Inhaled mainstream, exhaled mainstream, and
29 sidestream CS differ in composition. The CS contains two
30 classes of free radicals, one in the gas phase and another in
31 tar. The gas phase radicals consist of inorganic radicals (e.g.,
32 nitric oxide, NO) as well as organic radicals such as carbon-
33 and oxygen-centered radicals. Nitric oxide is slowly oxidized
34 to NO₂. It is estimated that there are approximately 10¹⁷
35 organic radicals per puff in gas phase smoke (Ref.46 and refer-
36 ence therein). Gas phase smoke is unstable and inactivates
37 ??1PI. In contrast, tar radicals in the particulate phase are
38 stable indefinitely and contain as many as 10¹⁸ free radicals
39

1 per gram, the major ones being quinone–hydroquinone com-
2 plex. This complex is an active redox system capable of redu-
3 cing molecular oxygen to produce superoxide, eventually
4 leading to H₂O₂ and OH radicals. Tar also chelates metals,
5 such as iron, that catalyze the decomposition of H₂O₂. An aqu-
6 eous suspension of tar produces hydroxyl radicals and has
7 been shown to cleave DNA. Many smokers have switched from
8 high- to low-tar cigarettes. Though low tar cigarettes may
9 expose the lungs to lower levels of carcinogens, they produce
10 a higher burden of oxidants. Nicotine is the most important
11 smoke component present in the blood of smokers, and it has
12 a half-life of 2 hr. Nicotine affects the respiratory, cardiovascu-
13 lar, central nervous, and the endocrine systems. Another
14 significant component of CS is Cd compounds, which have a
15 long half-life, accumulate in the lungs, and induce acute
16 inflammatory reactions in the lung and increased lung epithe-
17 lial permeability.

20 VI. WHAT IS NF- κ B?

21
22 The NF- κ B represents a group of five proteins namely c-Rel,
23 RelA (p65), Rel B, NF- κ B1 (p50 and p105), and NF- κ B2 (p52)
24 (16). The NF- κ B proteins are regulated by inhibitors of the
25 I κ B family, which includes I κ B α , I κ B β , I κ B ϵ , I κ B γ , Bcl-3,
26 p100, and p105 (47). In an inactive state, NF- κ B is present in
27 the cytoplasm as a heterotrimer consisting of p50, p65, and
28 I κ B α subunits. In response to an activation signal, the I κ B α
29 subunit is phosphorylated at serine residues 32 and 36, ubi-
30 quitinated at lysine residues 21 and 22 and degraded through
31 the proteosomal pathway, thus exposing the nuclear localiza-
32 tion signals on the p50-p65 heterodimer. The p65 is then
33 phosphorylated, leading to nuclear translocation and binding
34 to a specific sequence in DNA, which in turn results in gene
35 transcription. The phosphorylation of κ B α is catalyzed by
36 the IKK. The IKK consists of three subunits IKK- α , IKK- β ,
37 and IKK- γ (also called NEMO) (for references see Ref. 48).
38 Gene deletion studies have indicated that IKK- β is essential
39 for NF- κ B activation by most agents (49). The kinase that

1 induces the phosphorylation of p65 is controversial, but
2 IKK- β , protein kinase C, and protein kinase A have been
3 implicated (17–19).
4

5 **VII. RELEVANCE OF NF- κ B TO CIGARETTE** 6 **SMOKING** 7

8 There are several reasons to believe NF- κ B is a good target by AQ8
9 which to examine CS-induced lung cancer development and its
10 chemoprevention. First, benzo[a]pyrene, a component of CS,
11 has recently been shown to activate NF- κ B in lung adenocar-
12 cinoma cells (37) and in vascular smooth muscle cells (50).
13 Second, CS is also a potent source of ROIs (44–46), which are
14 required for NF- κ B activation (47). Our laboratory and others
15 have shown that antioxidants and overexpression of cells with
16 antioxidant enzymes such as Mn superoxide dismutase or with
17 γ -glutamylcysteinyl synthase (51–53) block NF- κ B activation.
18 Third, NF- κ B activation has been implicated in chemical car-
19 cinogenesis and tumorigenesis (54,55). Fourth, CS has been
20 shown to induce NF- κ B-regulated chemokine genes in bron-
21 chial epithelium (Ref. 38 and references therein). Lastly our
22 laboratory and others have shown that most chemopreventive
23 agents suppress NF- κ B activation (56–60).
24

25 **VIII. WHY NF- κ B IS IMPORTANT** 26 **FOR CANCER?** 27

28 The NF- κ B has been shown to regulate the expression of a
29 number of genes whose products are involved in tumorigen-
30 esis (20,21). These include antiapoptosis genes (e.g., cIAP, AQ9
31 survivin, TRAF, bcl-2, and bcl-xl) COX2; MMP-9; genes encod-
32 ing adhesion molecules, chemokines, inflammatory cytokines
33 and iNOS; and cell cycle regulatory genes (e.g., cyclin D1
34 (22)). Thus, agents that can suppress NF- κ B activation have
35 the potential to suppress carcinogenesis and have therapeutic
36 potential (21,23). The therapeutic role of phytochemicals in
37 prevention and treatment of cancer has been indicated
38 (24–26). Thus, plant-derived phytochemicals that could
39

1 suppress NF- κ B activation by various carcinogens have been
2 shown (Table 1). AQ10

3 4 5 **IX. CHEMOPREVENTIVE AGENTS INHIBIT** 6 **NF- κ B ACTIVATION**

7
8 Several agents that suppress carcinogenesis have been shown
9 to block NF- κ B activation. These include curcumin, green tea AQ11
10 polyphenols, silymarin, and resveratrol (Fig. 1). Curcumin is
11 a polyphenol (diferuloylmethane) derived from the roots of
12 *Curcuma longa*, and it inhibits both tumor initiation induced
13 by BP and 7,12 dimethylbenz(a)anthracene and phorbol ester-
14 induced tumor promotion (61–63). Both B[a]P and phorbol
15 esters are potent activators of NF- κ B. Curcumin has also been
16 shown to suppress the expression of several genes involved in
17 carcinogenesis including COX 2, lipooxygenases, and iNOS
18 (64–67), also known to require NF- κ B activation. Addition-
19 ally, our laboratory has shown that curcumin blocks the
20 TNF-induced expression of ICAM-1, VCAM-1, and ELAM-1,
21 all NF- κ B-regulated genes in endothelial cells, and needed
22 for tumor metastasis (68). Our laboratory has also shown that
23 curcumin suppresses the NF- κ B activation induced by var-
24 ious tumor promoters in different cell types (56). Similarly,
25 silymarin, derived from milk thistle (artichoks), has been AQ6
26 demonstrated to suppress carcinogenesis (69), and we have
27 shown that this compound also inhibits NF- κ B activation
28 through blocking the phosphorylation and degradation of
29 I κ B (59). Resveratrol, derived primarily from grapes and pea-
30 nuts, exhibits chemopreventive activity by inhibiting cellular
31 events associated with tumor initiation, promotion, and pro-
32 gression (70). Our laboratory and others showed that resver-
33 atrol also blocks NF- κ B activation and NF- κ B-regulated
34 expression of monocyte chemoattractant protein (MCP)-1
35 (60,71). Thus, several of these examples suggest that suppres-
36 sion of NF- κ B activation correlates with chemoprevention.

37 The epidemiological evidences also indicate that certain
38 cancers (e.g., breast, prostate, colon, and lung) are more
39 prevalent in the developed countries than in the developing

1 countries. It is most likely because of differences in dietary
2 constituents (16,17). We propose that there are constituents
3 of the every-day diet that regulate the activity of certain tran-
4 scription factors such as NF- κ B that plays a critical role in
5 carcinogenesis.

7 X. CONCLUSION

9 Evidence presented above suggests that activation of NF- κ B
10 can lead to tumor cell proliferation, invasion, angiogenesis,
11 and metastasis. Thus suppression of NF- κ B in cancer cells
12 may provide an additional target for prevention of cancer.
13 The NF- κ B blockers can also be considered for the therapy
14 of cancer, perhaps in combination with chemotherapeutic
15 agents or gamma irradiation.

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23 REFERENCES

- 24 1. Kelloff G IV. Perspectives on cancer-chemoprevention research
25 and drug development. *Adv Cancer Res* 2000; 78:199-334.
- 26 2. Aggarwal BB. Tumour necrosis factors receptor associated
27 signaling molecules and their role in activation of apoptosis,
28 JNK and NF-kappaB. *Ann Rheum Dis* 2000; 59(suppl 1):i6-16.
- 29 3. Komori A, Yatsunami J, Suganuma M, Okabe S, Abe S, Sakai A,
30 Sasaki K, Fujiki H. Tumor necrosis factor acts as a tumor
31 promoter in BALB/3T3 cell transformation. *Cancer Res* 1993;
32 53:1982-1985.
- 33 4. Suganuma M, Okabe S, Marino MW, Sakai A, Sueoka E,
34 Fujiki H. Essential role of tumor necrosis factor alpha (TNF-
35 alpha) in tumor promotion as revealed by TNF-alpha-deficient
36 mice. *Cancer Res* 1999; 59:4516-4518.
- 37
- 38
- 39

- 1 5. Orosz P, Echtenacher B, Falk W, Ruschoff J, Weber D, Mannel
2 DN. Enhancement of experimental metastasis by tumor necro-
3 sis factor. *J Exp Med* 1993; 177:1391–1398.
- 4 6. Hafner M, Orosz P, Kruger A, Mannel DN. TNF promotes metas-
5 tasis by impairing natural killer cell activity. *Int J Cancer* 1996;
6 66:388–392.
- 7 7. Orosz P, Kruger A, Hubbe M, Ruschoff J, Von Hoegen P,
8 Mannel DN. Promotion of experimental liver metastasis by
9 tumor necrosis factor. *Int J Cancer* 1995; 60:867–871.
- 10 8. Moore RJ, Owens DM, Stamp G, Arnott C, Burke F, East N,
11 Holdsworth H, Turner L, Rollins B, Pasparakis M, Kollias G,
12 Balkwill F. Mice deficient in tumor necrosis factor-alpha are
13 resistant to skin carcinogenesis. *Nat Med* 1999; 5:828–831.
- 14 9. Aggarwal BB, Schwarz L, Hogan ME, Rando RF. Triple helix-
15 forming oligodeoxyribonucleotides targeted to the human tumor
16 necrosis factor (TNF) gene inhibit TNF production and block the
17 TNF-dependent growth of human glioblastoma tumor cells.
18 *Cancer Res* 1996; 56:5156–5164.
- 19 10. Giri DK, Aggarwal BB. Constitutive activation of NF-kappaB
20 causes resistance to apoptosis in human cutaneous T cell
21 lymphoma HuT-78 cells. Autocrine role of tumor necrosis fac-
22 tor and reactive oxygen intermediates. *J Biol Chem* 1998; 273:
23 14008–14014.
- 24 11. Arnott CH, Scott KA, Moore RJ, Hewer A, Phillips DH, Parker P,
25 Balkwill FR, Owens DM. Tumour necrosis factor-alpha mediates
26 tumour promotion via a PKC alpha- and AP-1-dependent path-
27 way. *Oncogene* 2002; 22:4728–4738.
- 28 12. Suganuma M, Okabe S, Kurusu M, Iida N, Ohshima S, Saeki Y,
29 Kishimoto T, Fujiki H. Discrete roles of cytokines, TNF-alpha,
30 IL-1, IL-6 in tumor promotion and cell transformation. *Int*
31 *J Oncol* 2002; 20:131–136.
- 32 13. Hehlhans T, Stoelcker B, Stopfer P, Muller P, Cernaianu G,
33 Guba M, Steinbauer M, Nedospasov SA, Pfeffer K, Mannel
34 DN. Lymphotoxin-beta receptor immune interaction promotes
35 tumor growth by inducing angiogenesis. *Cancer Res* 2002; 62:
36 4034–4040.
- 37
38
39

- 1 14. Suganuma M, Okabe S, Sueoka E, Iida N, Komori A, Kim SJ,
2 Fujiki H. A new process of cancer prevention mediated through
3 inhibition of tumor necrosis factor alpha expression. *Cancer*
4 *Res* 1996; 56:3711–3715.
- 5 15. Sueoka N, Sueoka E, Okabe S, Fujiki H. Anti-cancer effects of
6 morphine through inhibition of tumour necrosis factor-alpha
7 release and mRNA expression. *Carcinogenesis* 1996; 17:
8 2337–2341.
- 9 16. Parkin DM. Cancer in developing countries. *Cancer Surv*
10 1994; 19–20:519–561.
- 11 17. Wingo PA, Ries LA, Giovino GA, Miller DS, Rosenberg HM,
12 Shopland DR, Thun MJ, Edwards BK. Annual report to the
13 nation on the status of cancer, 1973–1996, with a special
14 section on lung cancer and tobacco smoking. *J Natl Cancer*
15 *Inst* 1999; 1(8):675–690.
- 16 18. Stjernfeldt M, Ludvigsson J, Berglund K, Lindsten J. Mater-
17 nal smoking during pregnancy and the risk of childhood
18 cancer. *Lancet* 1986; 2(8508):687–688.
- 19 19. Sandier DP, Wilcox AJ, Everson RB. Cumulative effects of
20 lifetime passive smoking on cancer risk. *Lancet* 1985; 1(8424):
21 312–315.
- 22 20. Saracci R, Riboli E. Passive smoking and lung cancer: current
23 evidence and ongoing studies at the International Agency for
24 Research on Cancer. *Mutat Res* 1989; 222(2):117–127.
- 25 21. Bhisey RA, Ramchandani AG, D'Souza AV, Borges AM, Notani
26 PN. Long-term carcinogenicity of pan masala in Swiss mice.
27 *Int J Cancer* 1999; 83(5):679–684.
- 28 22. Hecht SS. Tobacco smoke carcinogens and lung cancer. *J Natl*
29 *Cancer Inst* 1999; 91(14):1194–1210.
- 30 23. Castonguay A, Pepin P, Stoner GD. Lung tumorigenicity of
31 NNK given orally to A/J mice: its application to chemopreven-
32 tive efficacy studies. *Exp Lung Res* 1991; 17(2):485–499.
- 33 24. Riddle MC, McDaniel PA. Acute reduction of renal 11 beta-
34 hydroxysteroid dehydrogenase activity by several antina-
35 triuretic stimuli. *Metab Clin Exp* 1993; 42(10):1370–1374.
- 36
37
38
39

- 1 25. Ahrendt SA I, Chow JT, Yang SC, Wu L, Zhang MJ, Jen J,
2 Sidransky D. Alcohol consumption and cigarette smoking
3 increase the frequency of p53 mutations in non-small cell lung
4 cancer. *Cancer Res* 2000; 60(12):3155–3159. AQ13
- 5 26. Kouri RE, McKinney CE, Slomiany DJ, Snodgrass DR, Wray NP,
6 McLemore TL. Positive correlation between high aryl hydrocar-
7 bon hydroxylase activity and primary lung cancer as analyzed
8 in cryopreserved lymphocytes. *Cancer Res* 1982; 42(12):
9 5030–5037.
- 10 27. McLemore TL, Martin RR, Toppell KL, Busbee DL, Cantrell ET.
11 Comparison of aryl hydrocarbon hydroxylase induction in
12 cultured blood lymphocytes and pulmonary macrophages. *J*
13 *Clin Invest* 1977; 60(5):1017–1024.
- 14 28. Korsgaard R, Trell E, Simonsson BG, Stiksa G, Janzon L,
15 Hood B, Oldbring J. Aryl hydrocarbon hydroxylase induction
16 levels in patients with malignant tumors associated with
17 smoking. *J Cancer Res Clin Oncol* 1984; 108(3):286–289.
- 18 29. Mace K, Bowman ED, Vautravers P, Shields PG, Harris CC,
19 Pfeifer AM. Characterisation of xenobiotic-metabolising enzyme
20 expression in human bronchial mucosa and peripheral lung
21 tissues. *Europ J Cancer* 1998; 34(6):914–920.
- 22 30. Cade JE, Margetts BM. Relationship between diet and smoking—
23 is the diet of smokers different? *J Epidemiol Community Health*
24 1991; 45(4):270–27.
- 25 31. Schlager JJ, Powis G. Cytosolic NAD(P)H:(quinone-acceptor)
26 oxidoreductase in human normal and tumor tissue: effects of
27 cigarette smoking and alcohol. *Intern J Cancer* 1990; 45:403–409.
- 28 32. Sinha BK, Katki AG, Batist G, Cowan KH, Myers CE.
29 Adriamycin-stimulated hydroxyl radical formation in human
30 breast tumor cells. *Biochem Pharmacol* 1987; 36(6):793–796.
- 31 33. Talcott RE, Levin VA. Glutathione-dependent denitrosation of
32 *N,N'*-bis(2-chloroethyl)*N*-nitrosourea (BCNU): nitrite release
33 catalyzed by mouse liver cytosol in vitro. *Drug Metab Disposi-*
34 *tion* 1983; 11(2):175–176.
- 35 34. Denissenko MF, Pao A, Tang M, Pfeifer GP. Preferential
36 formation of benzo[a]pyrene adducts at lung cancer mutational
37 hotspots in P53. *Science* 1996; 274(5286):430–432.
- 38
39

- 1 35. Hellin AC, Calmant P, Gielen J, Bours V, Merville MP. Nuclear factor—kappaB-dependent regulation of p53 gene
2 expression induced by daunomycin genotoxic drug. *Oncogene* AQ14
3 1998; 16(9):1187–1195.
4
- 5 36. Venkatachalam S, Denissenko M, Wani AA. Modulation of
6 (+/–)-anti-BPDE mediated p53 accumulation by inhibitors of
7 protein kinase C and poly(ADP-ribose) polymerase. *Oncogene*
8 1997; 14(7):801–809.
9
- 10 37. Pei XH, Nakanishi Y, Takayama K, Bai F, Hara N. Benzo[a]-
11 pyrene activates the human p53 gene through induction of
12 nuclear factor kappaB activity. *J Biol Chem* 1999; 274(49):
13 35240–35246.
14
- 15 38. Nishikawa M, Kakemizu N, Ito T, Kudo M, Kaneko T, Suzuki M,
16 Udaka N, Ikeda H, Okubo T. Superoxide mediates cigarette
17 smoke-induced infiltration of neutrophils into the airways
18 through nuclear factor-kappaB activation and IL-8 mRNA
19 expression in guinea pigs in vivo. *Am J Respir Cell Mol Biol*
20 1999; 20(2):189–198.
21
- 22 39. Repine JE, Bast A, Lankhorst L. Oxidative stress in chronic
23 obstructive pulmonary disease. *Oxidative Stress Study Group*
24 [Rev] [296 refs]. *Amer J Respir Crit Care Med* 1997; 156(2 Pt 1):
25 341–357.
26
- 27 40. Cantin A, Crystal RG. Oxidants, antioxidants and the patho-
28 genesis of emphysema [Rev] [77 Refs.]. *Euro J Respir Dis*
29 *Suppl* 1985; 139:7–17.
30
- 31 41. Nesje LA, Mjos OD. Plasma HDL cholesterol and the
32 subclasses HDL2 and HDL3 in smokers and non-smokers.
33 *Artery* 1985; 13(1):7–18.
34
- 35 42. Frei B, Forte TM, Ames BN, Cross CE. Gas phase oxidants of
36 cigarette smoke lipid induce peroxidation and changes in lipo-
37 protein properties in human blood plasma. Protective effects of
38 ascorbic acid. *Biochem J* 1991; 277(Pt 1):133–138.
39
- 34 43. Reznick AZ, Cross CE, Hu ML, Suzuki YJ, Khwaja S, Safadi A,
35 Motchnik PA, Packer L, Halliwell B. Modification of plasma
36 proteins by cigarette smoke as measured by protein carbonyl
37 formation. *Biochem J* 1992; 286(Pt 2):607–611.
38
39

- 1 44. Cross CE, O'Neill CA, Reznick AZ, Hu ML, Marcocci L, Packer L, Frei B. Cigarette smoke oxidation of human plasma constituents. *Ann NY Acad Sci* 1993; 686:72–89 [Discussion 89–90]. AQ15
- 2
- 3
- 4 45. Cosgrove JP, Borish ET, Church DF, Pryor WA. The metal-mediated formation of hydroxyl radical by aqueous extracts of cigarette tar. *Biochem Biophys Res Comm* 1985; 132:390–396.
- 5
- 6
- 7 46. Pryor WA. Cigarette smoke radicals and the role of free radicals in chemical carcinogenicity [Rev] [110 Refs.]. *Environ Health Perspect* 1997; 105(suppl 4):875–882.
- 8
- 9
- 10 47. Baeuerle PA, Baichwal VR. NF-kappa B as a frequent target for immunosuppressive and anti-inflammatory molecules [Rev] [105 Refs.]. *Adv Immunol* 1997; 65:111–137.
- 11
- 12
- 13 48. Karin M. The beginning of the end: IkappaB kinase (IKK) and NF- κ B activation. *J Biol Chem* 1999; 274(39):27339–27342.
- 14
- 15 49. Pahl HL. Activators and target genes of Rel/NF- κ B transcription factors. *Oncogene* 1999; 18:6853–6866.
- 16
- 17 50. Yan Z, Subbaramaiah K, Camilli T, Zhang F, Tanabe T, McCaffrey TA, Dannenberg AJ, Weksler BB. Benzo[a]pyrene induces the transcription of cyclooxygenase-2 in vascular smooth muscle cells. Evidence for the involvement of extracellular signal-regulated kinase and NF- κ B. *J Biol Chem* 2000; 275:4949–4955.
- 18
- 19
- 20
- 21
- 22
- 23 51. Manna S, Zhang HJ, Yan T, Oberley LW, Aggarwal BB. Overexpression of Mn-superoxide dismutase suppresses TNF-induced apoptosis and activation of nuclear transcription factor- κ B and activated protein-1. *J Biol Chem* 1998; 273:13245–13254.
- 24
- 25
- 26 52. Manna SK, Kuo MT, Aggarwal BB. Overexpression of γ -glutamylcysteine synthetase abolishes tumor necrosis factor-induced apoptosis and activation of nuclear transcription factor-kappa B and activator protein-1. *Oncogene* 1999; 18:4371–4382.
- 27
- 28
- 29
- 30 53. Schmidt KN, Amstad P, Cerutti P, Baeuerle PA. The roles of hydrogen peroxide and superoxide as messengers in the activation of transcription factor NF-kappa B. *Chemistry Biol* 1995; 2(1):13–22.
- 31
- 32
- 33 54. Sharma HW, Narayanan R. The NF- κ B transcription factor in oncogenesis. *Anticancer Res* 1996; 16:589–596.
- 34
- 35
- 36
- 37
- 38
- 39

- 1 55. Waddick KG, Uckun FM. Innovative treatment programs
2 against cancer: II. Nuclear factor-kappaB (NF-kappaB) as a
3 molecular target. *Biochem Pharmacol* 1999; 57:9–17.
- 4 56. Singh S, Aggarwal BB. Activation of transcription factor
5 NF- κ B is suppressed by curcumin (Diferulolylmethane). *J Biol*
6 *Chem* 1995; 270:24995–25000.
- 7 57. Natarajan K, Singh S, Burke TR Jr, Grunberger D, Aggarwal
8 BB. Caffeic acid phenethyl ester (CAPE) is a potent and speci-
9 fic inhibitor of activation of nuclear transcription factor
10 NF- κ B. *Proc Natl Acad Sci USA* 1996; 93:9090–9095.
- 11 58. Kumar K, Dhawan S, Aggarwal BB. Emodin (3-methyl-1,6,
12 8-trihydroxyanthraquinone) inhibits the TNF-induced NF- κ B
13 activation, I κ B degradation and expression cell surface adhe-
14 sion protein in human vascular endothelial cells. *Oncogene*
15 1998; 17:913–918.
- 16 59. Manna SK, Mukhopadhyay A, Van NT, Aggarwal BB. Sily-
17 marin suppresses TNF-induced activation of nuclear tran-
18 scription factor- κ B, c-Jun N-terminal kinase and apoptosis. *J*
19 *Immunol* 1999; 163:6800–6809.
- 20 60. Manna SK, Mukhopadhyay A, Aggarwal BB. Resveratrol
21 suppresses TNF-induced activation of nuclear transcription
22 factors NF- κ B, activator protein-1, and apoptosis: potential
23 role of reactive oxygen intermediates and lipid peroxidation.
24 *J Immunol* 2000; 164:6509–6519.
- 25 61. Huang MT, Wang ZY, Georgiadis CA, Laskin JD, Conney AH.
26 Inhibitory effects of curcumin on tumor initiation by benzo[a]-
27 pyrene and 7,12-dimethylbenz[a]anthracene. *Carcinogenesis*
28 1992; 13:2183–2186.
- 29 62. Huang MT, Smart RC, Wong C-Q, Conney AH. Inhibitory effect
30 of curcumin, chlorogenic acid, caffeic acid, and ferulic acid on
31 tumor promotion in mouse skin by 12-*O*-tetradecanoylphorbol-
32 13-acetate. *Cancer Res* 1988; 48:5941–5946.
- 33 63. Conney AM, Lysz T, Ferraro T, Abidi TF, Manchand PS,
34 Laskin JD, Huang MT. Inhibitory effect of curcumin and
35 some related dietary compounds on tumor promotion and
36 arachidonic acid metabolism in mouse skin. *Adv Enzyme Regul*
37 1991; 31:385–396.
- 38
- 39

- 1 64. Plummer SM, Holloway KA, Manson MM, Munks RJ, Kaptein A,
2 Farrow S, Howells L. Inhibition of cyclo-oxygenase 2 expression in
3 colon cells by the chemopreventive agent curcumin involves
4 inhibition of NF-kappaB activation via the NIK/IKK signalling
5 complex. *Oncogene* 1999; 18(44):6013–6020.
- 6 65. Zhang F, Altorki NK, Mestre JR, Subbaramaiah K, Dannen-
7 berg AJ. Curcumin inhibits cyclooxygenase-2 transcription in
8 bile acid- and phorbol ester-treated human gastrointestinal
9 epithelial cells. *Carcinogenesis* 1999; 20(3):445–451.
- 10 66. Began G, Sudharshan E, Appu Rao AG. Inhibition of lipoxygen-
11 ase 1 by phosphatidylcholine micelles-bound curcumin. *Lipids*
12 1998; 33(12):1223–1228.
- 13 67. Chan MM, Huang HI, Fenton MR, Fong D. In vivo inhibition
14 of nitric oxide synthase gene expression by curcumin, a cancer
15 preventive natural product with anti-inflammatory properties.
16 *Biochem Pharmacol* 1998; 55:1955–1962.
- 17 68. Kumar A, Dhawan S, Hardegen NJ, Aggarwal BB. Curcumin
18 (Diferuloylmethane) inhibition of tumor necrosis factor (TNF)-
19 mediated adhesion of monocytes to endothelial cells by
20 suppression of cell surface expression of adhesion molecules
21 and of nuclear factor-kappaB activation. *Biochem Pharmacol*
22 1998; 55(6):775–783.
- 23 69. Lahiri-Chatterjee M, Katiyar SK, Mohari RR, Agarwal R. A
24 flavonoid antioxidant, silymarin, affords exceptionally high
25 protection against tumor promotion in the SENCAR mouse
26 skin tumorigenesis model. *Cancer Res* 1999; 59:622–632.
- 27 70. Jang M, Cai L, Udeani GO, Slowing KV, Thomas CF, Beecher CW,
28 Fong HH, Farnsworth NR, Kinghorn AD, Mehta RG, Moon RC,
29 Pezzuto JM. Cancer chemopreventive activity of resveratrol, a
30 natural product derived from grapes. *Science* 1997:275–218.
- 31 71. Holmes-McNary M, Baldwin AS Jr. Chemopreventive proper-
32 ties of trans-resveratrol are associated with inhibition of
33 activation of the IkappaB kinase. *Cancer Res* 2000; 60(13):
34 3477–3483.
- 35
36
37
38
39